

Research article

Open Access

Molecular evolution of *Phox*-related regulatory subunits for NADPH oxidase enzymes

Tsukasa Kawahara* and J David Lambeth

Address: Department of Pathology and Laboratory Medicine, Emory University School of Medicine, Atlanta, Georgia, 30322, USA

Email: Tsukasa Kawahara* - tkawaha@emory.edu; J David Lambeth - noxdoc@mac.com

* Corresponding author

Published: 27 September 2007

Received: 16 June 2007

BMC Evolutionary Biology 2007, 7:178 doi:10.1186/1471-2148-7-178

Accepted: 27 September 2007

This article is available from: <http://www.biomedcentral.com/1471-2148/7/178>

© 2007 Kawahara and Lambeth; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: The reactive oxygen-generating NADPH oxidases (Noxes) function in a variety of biological roles, and can be broadly classified into those that are regulated by subunit interactions and those that are regulated by calcium. The prototypical subunit-regulated Nox, Nox2, is the membrane-associated catalytic subunit of the phagocyte NADPH-oxidase. Nox2 forms a heterodimer with the integral membrane protein, p22*phox*, and this heterodimer binds to the regulatory subunits p47*phox*, p67*phox*, p40*phox* and the small GTPase Rac, triggering superoxide generation. Nox-organizer protein 1 (NOXO1) and Nox-activator 1 (NOXA1), respective homologs of p47*phox* and p67*phox*, together with p22*phox* and Rac, activate Nox1, a non-phagocytic homolog of Nox2. NOXO1 and p22*phox* also regulate Nox3, whereas Nox4 requires only p22*phox*. In this study, we have assembled and analyzed amino acid sequences of Nox regulatory subunit orthologs from vertebrates, a urochordate, an echinoderm, a mollusc, a cnidarian, a choanoflagellate, fungi and a slime mold amoeba to investigate the evolutionary history of these subunits.

Results: Ancestral p47*phox*, p67*phox*, and p22*phox* genes are broadly seen in the metazoa, except for the ecdysozoans. The choanoflagellate *Monosiga brevicollis*, the unicellular organism that is the closest relatives of multicellular animals, encodes early prototypes of p22*phox*, p47*phox* as well as the earliest known Nox2-like ancestor of the Nox1-3 subfamily. p67*phox*- and p47*phox*-like genes are seen in the sea urchin *Strongylocentrotus purpuratus* and the limpet *Lottia gigantea* that also possess Nox2-like co-orthologs of vertebrate Nox1-3. Duplication of primordial p47*phox* and p67*phox* genes occurred in vertebrates, with the duplicated branches evolving into NOXO1 and NOXA1. Analysis of characteristic domains of regulatory subunits suggests a novel view of the evolution of Nox: in fish, p40*phox* participated in regulating both Nox1 and Nox2, but after the appearance of mammals, Nox1 (but not Nox2) became independent of p40*phox*. In the fish *Oryzias latipes*, a NOXO1 ortholog retains an autoinhibitory region that is characteristic of mammalian p47*phox*, and this was subsequently lost from NOXO1 in later vertebrates. Detailed amino acid sequence comparisons identified both putative key residues conserved in characteristic domains and previously unidentified conserved regions. Also, candidate organizer/activator proteins in fungi and amoeba are identified and hypothetical activation models are suggested.

Conclusion: This is the first report to provide the comprehensive view of the molecular evolution of regulatory subunits for Nox enzymes. This approach provides clues for understanding the evolution of biochemical and physiological functions for regulatory-subunit-dependent Nox enzymes.

Background

Nox enzymes (reactive oxygen-generating NADPH-oxidases) diverged early in evolution into calcium-regulated Noxes (e.g., Nox5 and the Duox enzymes) and Noxes that are activated by binding to regulatory subunits [1,2]. The most extensively studied of the latter is the phagocyte NADPH-oxidase (*Phox*), whose role in host defense has been documented at length [3-8]. Professional phagocytes such as neutrophils and macrophages produce large amounts of superoxide, with secondary production of other microbicidal reactive oxygen species (ROS). Superoxide is generated by the *Phox*, which consists of the catalytic subunit gp91*phox* (a.k.a. Nox2), along with the regulatory subunits p22*phox*, p67*phox*, p47*phox*, p40*phox*, and the small GTPase, Rac [3,6,9-11]. The importance of the oxidase is demonstrated by the inherited condition, chronic granulomatous disease (CGD), in which absent or mutated *Phox* proteins result in an inability of phagocytes to kill microbes [4,12,13].

Nox2 and p22*phox* are integral membrane proteins that form a heterodimer referred to as flavocytochrome *b*₅₅₈. In resting cells, flavocytochrome *b*₅₅₈ is inactive and p47*phox*, p67*phox*, and p40*phox* are all present in the cytoplasm. Cell activation is accompanied by phosphorylation of p47*phox* and probably other *Phox*-regulatory subunits [3,10,14-16], and by guanine nucleotide exchange factors that convert Rac-GDP to Rac-GTP [3,17]. These events trigger assembly of these proteins at the membrane, resulting in activation of Nox2. An intricate set of protein-protein interactions facilitates the binding of the regulatory subunits to the membrane components and to each other, and these interactions are mediated by well-characterized modular interaction domains. For example, p47*phox* and p67*phox* bind via a C-terminal proline-rich region (PRR) in p47*phox* and a C-terminal Src homology 3 (SH3) domain in p67*phox* [18]. In the non-activated cell, the p47*phox*-p67*phox* complex fails to bind to flavocytochrome *b*₅₅₈, due to an unusual autoinhibitory mechanism. In the resting cell, tandem SH3 domains in p47*phox* (referred to as the *bis*-SH3 domain) bind to an auto-inhibitory region (AIR), preventing the *bis*-SH3 domain from binding to the PRR of p22*phox*. Upon cell activation, serine residues of the AIR become phosphorylated, releasing the *bis*-SH3 domain so that it can now bind to p22*phox* [19-22]. In addition, p47*phox* possesses another membrane-binding region, the PX domain, which binds to the headgroups of phosphatidylinositols present in the membrane [23-25]. Together, these interactions with membrane lipids and p22*phox* promote the assembly of p47*phox* and p67*phox* with flavocytochrome *b*₅₅₈. Concurrently, Rac-GTP translocates to the membrane where it interacts with the N-terminal tetratricopeptide repeat (TPR) region of p67*phox* [26-28]. An activation domain (AD) in p67*phox*, which is conformationally reg-

ulated by Rac binding [29], activates the hydride transfer from NADPH to FAD in Nox2. Subsequent electron transfer through two heme groups permits reduction of oxygen to form superoxide [30,31]. p40*phox* binds through its *Phox*/Bem 1 (PB1) domain to a partner PB1 domain in p67*phox* and facilitates the assembly of p67*phox*-p47*phox* at the membrane through lipid binding via its PX domain [32-34]. Although no CGD patients have been described with mutations in p40*phox*, neutrophils from p40*phox*-knockout mice exhibit CGD-like severe defects in Nox2-derived oxidant-dependent bacterial killing [35], also suggesting that p40*phox* is a crucial component for Nox2 function.

The first homologue of gp91*phox* identified was Nox1, which is abundantly expressed in non-phagocyte cells including gastrointestinal epithelium and vascular smooth muscle [9,36]. Subsequently, additional homologues were identified, and the Nox/Duox family in humans now consists of seven members: Nox1 though Nox5, Duox1 and Duox2 [9,37]. All members of Nox/Duox family contain the six transmembrane α -helical heme-binding domain and an FAD domain containing an NADPH binding site, which together constitute the "Nox domain" [1]. Nox5 and Duox1/2 contain a calcium-binding EF-hand motif. The evolution of these Noxes and their calcium-binding domains were considered in the recent article [1] and are not discussed here. The remaining Noxes, Nox1-4, require p22*phox* for activity, and Nox1-3 all require additional *Phox*-like regulatory subunits, reviewed recently [10,11,37].

Nox1, like gp91*phox*, is activated by regulatory subunits: NOXO1 (Nox-organizer protein 1) is a homologue of p47*phox* and NOXA1 (Nox-activator protein 1) is a homologue of p67*phox* [38-41]. Recent studies have also demonstrated activation of the Nox1 system by Rac1 [42-45]. NOXO1 and NOXA1 are co-expressed with Nox1 in colon epithelium and gastric mucosa [39,43,46], consistent with *in vivo* regulation of Nox1 by these novel subunits. Activation of Nox1 by regulatory subunits differs in two major respects compared to gp91*phox*. First, NOXO1 lacks the AIR and its associated regulatory phosphorylation sites that are present in p47*phox*. Consistent with the domain structure, NOXO1 co-localizes with Nox1/p22*phox* complex constitutively at/near the plasma membrane *in vivo* [40,41]. On the other hand, a recent study revealed that the *bis*-SH3 domain of NOXO1 bind to the extremely C-terminal PRR of NOXO1 [47]. Phosphorylation-independent disruption of the intramolecular interaction facilitates NOXO1 binding to the PRR of p22*phox*. Second, NOXA1 lacks critical basic amino acid residues in its PB1 domain that are important for binding to p40*phox*. NOXA1 does not interact with p40*phox*, making human Nox1 activity independent of p40*phox* [40]. Furthermore,

human NOXA1 lacks the central SH3 domain that is present in p67phox [38-40], but whose function is unknown. Like Nox1, Nox3 is also regulated by p22phox and NOXO1, but in humans does not require NOXA1 [48-51]. In comparison, Nox4 requires p22phox for activity, but other known regulatory subunits are not needed [52-54].

Another subfamily of the Noxes is the NoxA/NoxB group, found in a slime mold amoeba and fungi. These fungal Nox genes have been identified in most ascomycetes except for hemiascomycete yeasts, and also in higher basidiomycetes and chytridiomycetes (reviewed in [55]). No Nox gene has been identified in fungi that belong to Zygomycota and Micropordia. These NoxA/NoxB genes are closely related to the subunit-regulated Noxes according to taxonomy [1]. A non-animal homologue of p67phox/NOXA1 was described in slime mold amoeba *Dictyostelium discoideum* (*D. discoideum*) [56]. The p67phox homolog (referred to here as the Dd-p67-like) of *D. discoideum* possesses the N-terminal TPR region and PB1 domain characteristic of p67phox, but lacks the SH3 domain [56]. A p67phox homolog Nox-regulator (NOXR) was also reported in some fungi including ascomycetes and higher basidiomycetes [55,57]. The basic motif structure of NOXR is similar to that of *D. discoideum*. A NOXR-deficient mutant of the ascomycete *Epichloe festucae* shows decreased ROS-generation [57]. Taken together, these data suggest that NOXR regulates NoxA and/or NoxB in the slime mold and fungi, although this has not been tested directly.

Herein, we assembled and analyzed experimentally cloned and computationally predicted amino acid sequences of the Phox-related regulatory subunits p47phox, NOXO1, p67phox, NOXA1, p40phox, p22phox, and NOXR from 10 vertebrates, one urochordate, one echinoderm, one mollusc, one cnidarian, one choanoflagellate, 13 fungi, and one slime mold amoeba. Using this large set of amino acid sequences, we report: (i) the occurrence and origin of Phox-like subunits during evolution, (ii) identification of putative key amino acid residues conserved in regulatory subunits, (iii) the description of putative new modular regions that are conserved among Phox-related regulatory subunits, (iv) the molecular evolution of characteristic domains of the Phox-related regulatory subunits, and (v) a prediction of novel fungal and amoebal Nox organizer/enhancer subunits based on sequence analysis.

Results and discussion

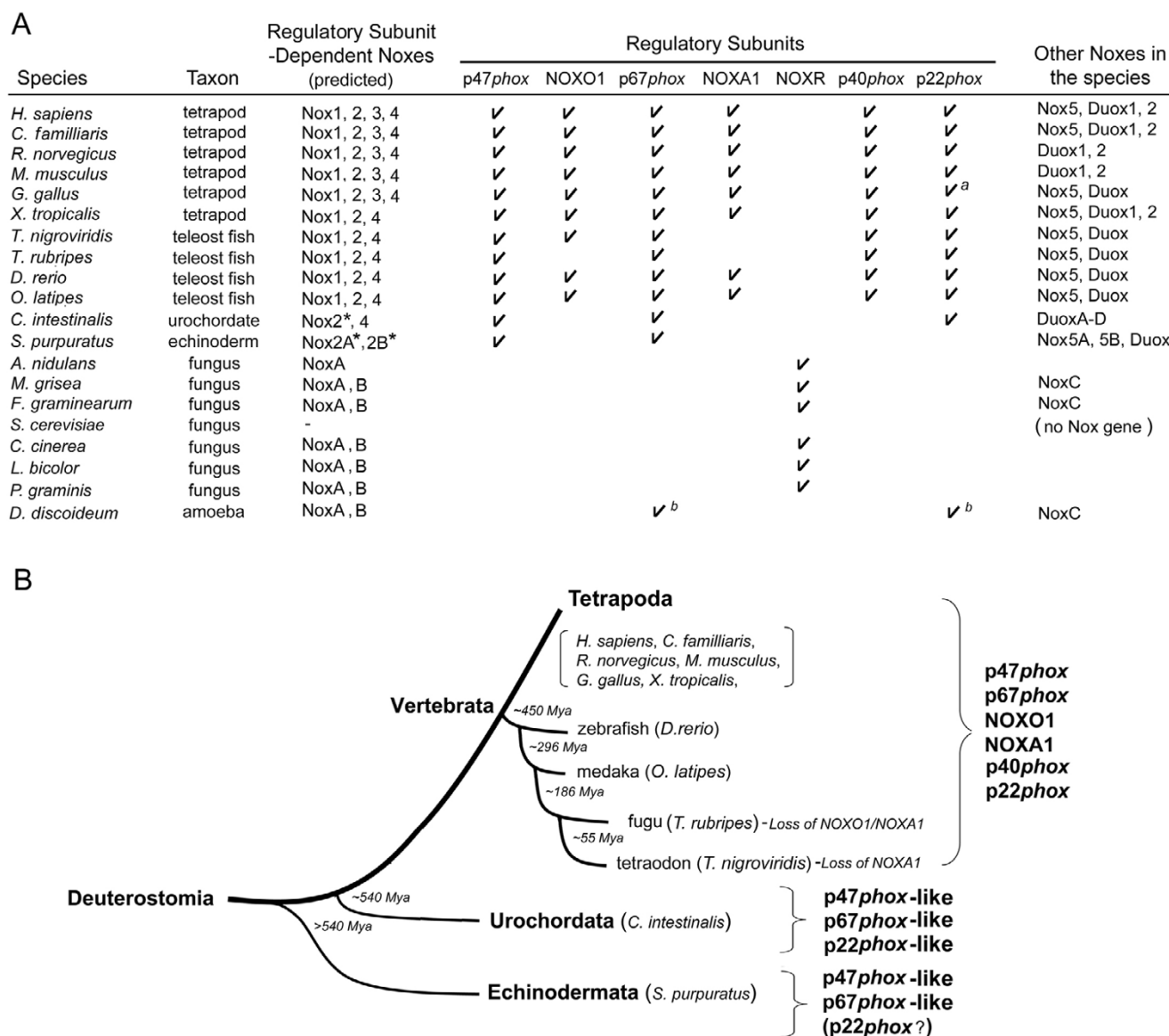
Occurrence of Phox-related regulatory subunits

Figure 1A summarizes the occurrence of subunit-dependent-like Nox orthologs along with the Phox-like regulatory subunit orthologs in the genomes of deuterostomes, fungi,

a slime mold amoeba. The tetrapods [human *Homo sapiens* (*H. sapiens*), dog *Canis familiaris* (*C. familiaris*), rat *Rattus norvegicus* (*R. norvegicus*), mouse *Mus musculus* (*M. musculus*), chicken *Gallus gallus* (*G. gallus*), frog *Xenopus tropicalis* (*X. tropicalis*)], and the two teleost fish [zebrafish *Danio rerio* (*D. rerio*), medaka *Oryzias latipes* (*O. latipes*)] have two isoforms of Nox organizer proteins (p47phox and NOXO1), two isoforms of Nox activator proteins (p67phox and NOXA1), and single isoforms of p40phox and p22phox (Figure 1A, the sequences are provided in Additional file 1). The pufferfish *Takifugu rubripes* (*T. rubripes*), which has a compact genome size, ~400 megabase (Mb) [58], lacks NOXO1 and NOXA1 although it possesses both Nox1 and Nox2 as well as p22phox, p47phox, p67phox, and p40phox (Figure 1A), in contrast to *D. rerio* and *O. latipes* that have less compact genomes (1680–2280 Mb and 870–1100 Mb, respectively, according to the Animal Genome Size Database [59]). Interestingly, *Tetraodon nigroviridis* (*T. nigroviridis*), another species of pufferfish, which also possesses a compact genome, ~340 Mb [60], encodes a NOXO1 ortholog, but not NOXA1 (Figure 1A).

The earlier chordate *Ciona intestinalis* (*C. intestinalis*) possesses a Nox2 ortholog, but not Nox1 or Nox3 [1,61]. The sea urchin *Strongylocentrotus purpuratus* (*S. purpuratus*) belongs to Echinodermata, which diverged early from an ancestor common to urochordates and vertebrates [62]. *S. purpuratus*, possesses two Nox2 genes, Nox2A and Nox2B, and an analysis of the taxonomy Noxes demonstrates that these Nox2 genes are related to the common ancestor gene of vertebrate Nox1, Nox2 and Nox3 [1]. *C. intestinalis* possesses single copies of p47phox-, p67phox-, and p22phox-like genes [61], which presumably regulate its Nox2 ortholog (Figure 1A). The *S. purpuratus* genome possesses single copies of p47phox- and p67phox-like genes (Figure 1A). No p22phox ortholog of *S. purpuratus* was identified using a BLAST search of recently published genome sequences [63]. Because the *S. purpuratus* p47phox-like protein has a highly conserved tandem SH3 region that in other species binds to the C-terminal proline-rich region (PRR) of p22phox, it seems likely that *S. purpuratus* has an as-yet unidentified p22phox homologue.

Using the estimated time of evolution of deuterostomes based on Hox gene clusters to construct a phylogenetic tree [64], we illustrate the occurrence during evolution of the Phox-related regulatory subunits (Figure 1B). The phylogenetic tree suggests an evolutionary model in which duplication of both the p47phox gene and the p67phox gene occurred at a time corresponding to the appearance of vertebrates, with the NOXO1 and NOXA1 genes subsequently evolving as distinct regulatory subunits in vertebrates (Figure 1B). This appearance of NOXO1 and NOXA1 coincided with the emergence of Nox1 [1]. The pufferfish *T. rubripes* and *T. nigroviridis* are considered to

**Figure 1****Relationships between the phylogenetic tree of species and the occurrence of Phox-related regulatory subunits.**

(A) Summary of the occurrence of Nox-regulatory subunit genes in vertebrates, a urochordate, an echinoderm, fungi, and a slime mold amoeba are shown. Asterisks indicate that the gene is a co-ortholog ancestor of all members of the vertebrate Nox1-3 subgroup. Superscripted letters represent: (a), a partial amino acid sequence predicted from genomic DNA fragment (nucleotides 713622–713565 of GenBank™ No. [NW_001471438.1](#)), (b) the gene taxonomy is discussed in the text. (B) Summary of the occurrence of Nox-regulatory subunit genes within Deuterostomia. The number in the fork of each branch indicates the estimated time in millions of years (Mya, millions of years ago) since a given species diverged. A schematic phylogeny and the estimated time of occurrence of species were created from genomic information [62, 125]. Branch lengths are for illustrative purposes and are not proportional to time since divergence.

have evolved from an *O. latipes* ancestor approximately 186 million years ago [64]. This tree suggests that NOXA1 and NOXO1 of *T. rubripes* and NOXA1 of *T. nigroviridis* have been lost during the decrease in genome sizes in

these species (Figure 1B). In contrast to the co-appearance of p47phox and p67phox orthologs with the earliest Nox2 ortholog in *S. purpuratus*, p40phox appeared later, in vertebrates, and was not found in the genomes of either *C.*

intestinalis or *S. purpuratus* (Figure 1B). The situation of the regulatory subunits of other primitive animals and a choanoflagellate is described below.

A homologue of p67phox/NOXA1 occurs in the slime mold amoeba (*D. discoideum*) [56] (Figure 1A). Fungal genomes of *Aspergillus nidulans* (*A. nidulans*), *Magnaporthe grisea* (*M. grisea*), and *Fusarium graminearum* (*F. graminearum*), all members of Ascomycota, also possess NOXR, a homolog of p67phox [55] (Figure 1A). However, *Saccharomyces cerevisiae* (*S. cerevisiae*) that also belongs to Ascomycota does not possess any Nox or NOXR orthologs [1,55] (Figure 1A). Some basidiomycetes [*Coprinopsis cinerea* (*C. cinerea*), *Laccaria bicolor* (*L. bicolor*), and *Postia placenta* (*P. placenta*)] also possess NOXR and NoxA/B genes (Figure 1A). The situation of the regulatory subunits of other fungi is described below. Interestingly, the fungi seem to lack precise homologues of p47phox/NOXO1 or its binding partner p22phox (Figure 1A). Recently, the possibility of p22phox-like gene in the *D. discoideum* genome was suggested [56], but it is not clear whether the gene represents an ancestral p22phox, a later adaptation or a functionally unrelated protein. We discuss the possibilities below based on a comparison of the amino acid sequences of amoebal p22phox and other p22phox orthologs. In addition, we identified other predicted proteins encoded in fungal and amoebal genomes that may function as (a) binding partner(s) of the Nox activator proteins (described below), suggesting the early evolutionary origin not only of the activator protein, but also possibly of a rudimentary organizer subunit.

Synten of NOXO1 and NOXA1 genes in vertebrates

An extensive BLAST search has failed to identify the NOXA1 gene in the pufferfish *T. rubripes* and *T. nigroviridis* genomes, as indicated in Figure 1. In addition to the absence of NOXA1, *T. rubripes* seems to have lost the NOXO1 gene. To substantiate that the NOXO1 and NOXA1 genes were indeed absent, we compared chromosomal synteny (the preserved chromosomal order of genes among organisms) of NOXO1 in vertebrates. The human NOXO1 gene is preceded by SYNGR3 and GFER genes on chromosome 16 (Figure 2A, abbreviations of gene names are shown in Additional file 2), and followed by TBL3, RPS2 and RPL3L. Synteny was preserved in mammals and the chicken *G. gallus*. The frog *X. tropicalis* lacks SYNGR3, but other markers are preserved. Among fish, *T. nigroviridis* conserved part of the mammalian synteny: NOXO1, TBL3, RPS2, and RPL3L and instead of SYNGR3 and GFER, C17orf39 is positioned next to NOXO1. Synteny of the *T. rubripes* genome was very similar to that of *T. nigroviridis*, although it lacks a NOXO1 ortholog (Figure 2A). Two fishes, *D. rerio* and *O. latipes*, genomes possess NOXO1 genes, but with greater variations in synteny; although TBL3 and C17orf39 were

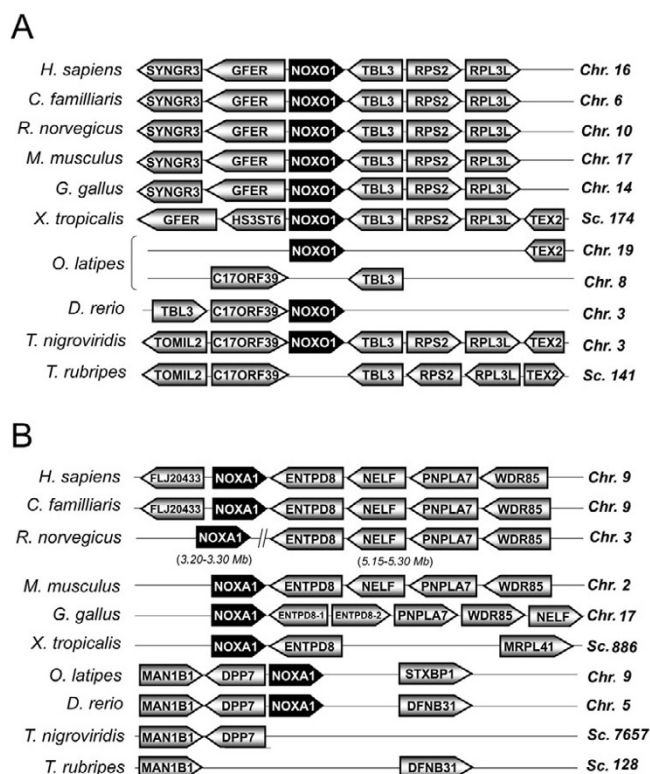


Figure 2

Syntenies of NOXO1 and NOXA1 genes. Syntenies of the indicated vertebrate NOXO1 (A) and NOXA1 (B) genes are shown. Genes are aligned in columns to illustrate orthology. Chromosome (Chr) or scaffold (Sc) number is indicated on the right. Abbreviations of gene names are shown in Additional file 2.

present in the same chromosome as NOXO1, other linked markers were not seen (Figure 2A). The gene markers C17orf39 and TBL3 of *O. latipes* are present on a separate chromosome from that of NOXO1. Thus, the synteny analysis of the *T. rubripes* genomes shows that the NOXO1 gene was indeed lost during evolution.

The synteny of genetic markers surrounding NOXA1 was highly conserved in *H. sapiens*, *C. familiaris*, *M. musculus*, *G. gallus* (Figure 2B). Both NOXA1 and the linked markers of *R. norvegicus* were present in chromosome 3, with a long insert between NOXA1 and ENTPD8 (Figure 2B). In the genome of *X. tropicalis*, synteny was more varied in that NOXA1 and ENTPD8 were present in the same scaffold, but other markers were not seen. Synteny of fish NOXA1 was preserved among the class but different from other vertebrates. *O. latipes* and *D. rerio* NOXA1 genes are preceded by MAN1B1 and DPP7 on chromosome 9 and 5, respectively (Figure 2B). *T. nigroviridis* possesses two

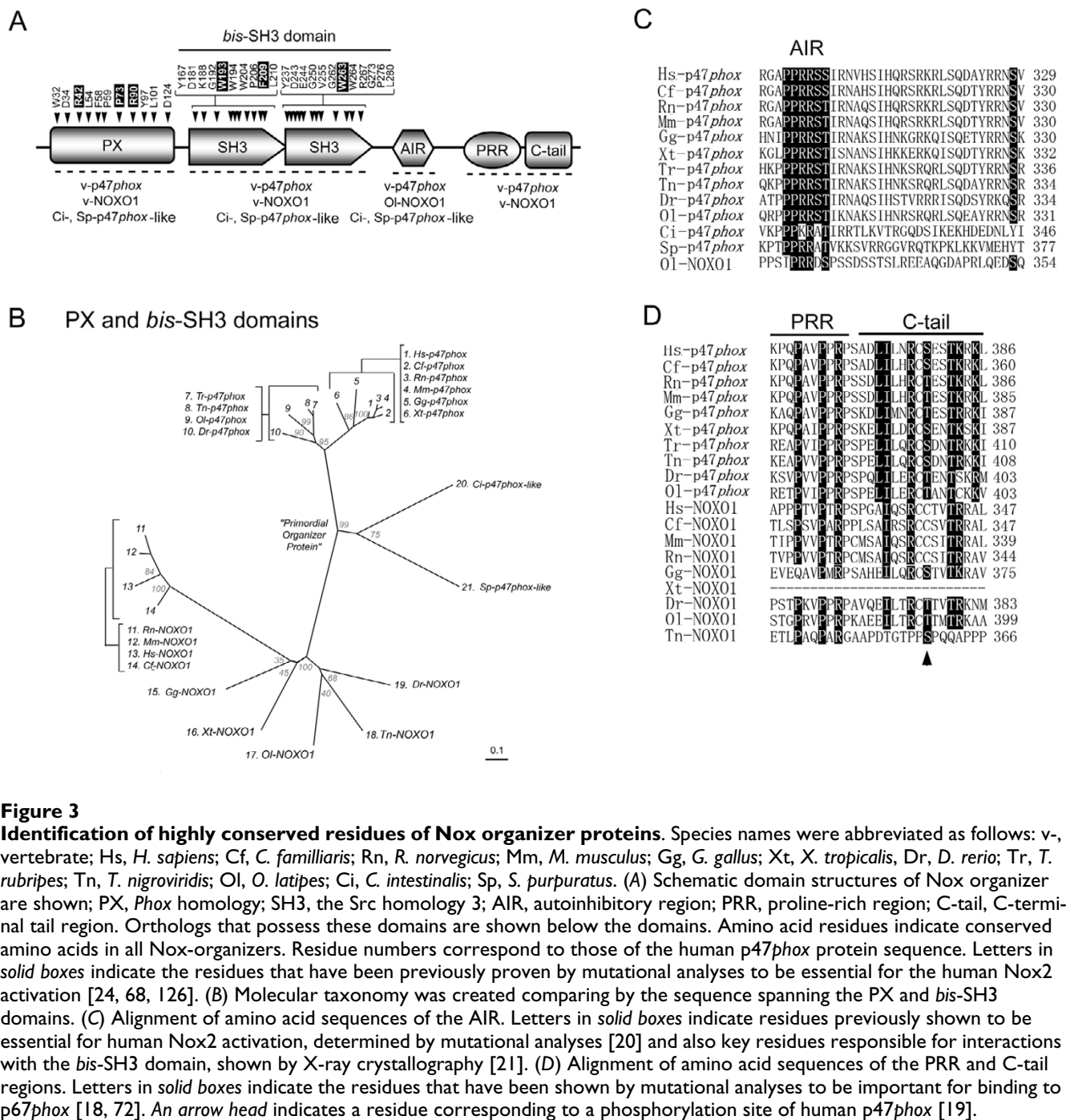


Figure 3
Identification of highly conserved residues of Nox organizer proteins. Species names were abbreviated as follows: v-, vertebrate; Hs, *H. sapiens*; Cf, *C. familiaris*; Rn, *R. norvegicus*; Mm, *M. musculus*; Gg, *G. gallus*; Xt, *X. tropicalis*; Dr, *D. rerio*; Tr, *T. rubripes*; Tn, *T. nigroviridis*; Ol, *O. latipes*; Ci, *C. intestinalis*; Sp, *S. purpuratus*. (A) Schematic domain structures of Nox organizer are shown; PX, Phox homology; SH3, the Src homology 3; AIR, autoinhibitory region; PRR, proline-rich region; C-tail, C-terminal tail region. Orthologs that possess these domains are shown below the domains. Amino acid residues indicate conserved amino acids in all Nox-organizers. Residue numbers correspond to those of the human p47phox protein sequence. Letters in solid boxes indicate the residues that have been previously proven by mutational analyses to be essential for the human Nox2 activation [24, 68, 126]. (B) Molecular taxonomy was created comparing by the sequence spanning the PX and bis-SH3 domains. (C) Alignment of amino acid sequences of the AIR. Letters in solid boxes indicate residues previously shown to be essential for human Nox2 activation, determined by mutational analyses [20] and also key residues responsible for interactions with the bis-SH3 domain, shown by X-ray crystallography [21]. (D) Alignment of amino acid sequences of the PRR and C-tail regions. Letters in solid boxes indicate the residues that have been shown by mutational analyses to be important for binding to p67phox [18, 72]. An arrow head indicates a residue corresponding to a phosphorylation site of human p47phox [19].

markers MAN1B1 and DPP7, but lacks the NOXA1 ortholog. Synteny of *T. rubripes* is similar to that of *D. rerio*, but a NOXA1 ortholog is absent. Thus, the synteny of NOXA1 had considerable variation, and this analysis indicates that NOXA1 orthologs were lost during the evolution of the two pufferfish genomes.

Identification of putative critical amino acids conserved among p47phox/NOXO1 orthologs
The PX domain of p47phox binds with highest affinity to phosphatidylinositol 3,4-bisphosphate [PtdIns(3,4)P₂], and with lower but significant affinity to PtdIns(3,4,5)P₃, PtdIns(3,5)P₂ and PtdIns(3)P [23-25]. The PX domain of

human NOXO1 shows a different lipid binding specificity, with highest affinity for PtdIns(3,5)P₂, PtdIns(5)P, and PtdIns(4)P and lower affinity for PtdIns(3)P and PtdIns(3,4)P₂ [41,65]. Both PX and *bis*-SH3 domains are seen in all p47phox/NOXO1 orthologs of deuterostomes (Figure 3A). Comparison of the sequences of the organizer proteins allowed identification of amino acid residues that are highly conserved (Figure 3A). Eleven highly conserved residues were identified in the PX domain, and 22 in the *bis*-SH3 domains (arrows in Figure 3A). Arg-42, Pro-73, and Arg-90 of the PX-domain and Trp-193, Phe-209, and Trp-263 of the *bis*-SH3 domain (human-p47phox numbering, black boxes in Figure 3A) were previously shown by mutational analyses to be essential for the activation of human Nox2 by p47phox [24,66-68].

While the PX and *bis*-SH3 domains are shared among all p47phox and NOXO1 orthologs including those of *S. purpuratus* and *C. intestinalis*, the molecular taxonomy of the regions (excluding the variable C-terminal regulator docking regions) reveals that vertebrate p47phox and NOXO1 families form distinct subgroups (Figure 3B), likely accounting, for example, for differences in lipid binding specificities [24,41]. The phylogenetic tree supports an evolutionary model in which the p47phox-like proteins of *S. purpuratus* and *C. intestinalis* (#21 and 20 of Figure 3B, respectively) branch from a root that is closely related to the primordial ancestors of both p47phox (#1-10 of Figure 3B) and NOXO1 (#11-19 of Figure 3B).

To maintain p47phox strictly in an inactive state that cannot bind to p22phox, human p47phox masks its *bis*-SH3 domain with the AIR region [21,68,69], and this region is absent in human NOXO1 [38-41]. Therefore Nox2 activity is considered more tightly controlled than Nox1 activity. Most residues essential for the function of the AIR (e.g., Pro-299, Pro-300, Arg-301, Arg-302, Ser-303, Ser-304 and Ser-328 of human p47phox), are well conserved among all vertebrate p47phox proteins (black boxes of Figure 3C). Interestingly *C. intestinalis* and *S. purpuratus* p47phox proteins possessed the AIR-like region (Figure 3C). *S. purpuratus* possesses two Nox2-like proteins Nox2A and 2B that are co-orthologs of vertebrate Noxes 1-3; the presence of a p47phox ortholog harboring the AIR suggests that this primitive Nox system functions analogously to the mammalian Nox2 system. All NOXO1 orthologs lack the AIR, with one exception, the teleost fish *O. latipes*, which retains the predicted AIR in both the p47phox and the NOXO1 genes (Figure 3C). Thus, this analysis supports an evolutionary model in which the NOXO1 "AIR-independent" Nox1 and Nox3 activation systems originated from the p47phox-like "AIR-dependent" Nox2 regulatory system. The *O. latipes* NOXO1, which contains the AIR, represents an evolutionary rem-

nant reflecting the transition from AIR-dependence to -independence.

The heterodimerization of human p47phox and p67phox is dependent on the interaction between the C-terminal PRR of p47phox and the C-terminal SH3 domain of p67phox [40,70-72]. Both of these regions were conserved in vertebrate p47phox and p67phox orthologs, but not in orthologs of lower organisms (Figure 3A). The alignment of vertebrate p47phox and NOXO1 amino acid sequences demonstrates that in addition to the PRR, the C-terminal region immediately following the PRR ("C-tail" in Figure 3D) was also highly conserved. The PRR of p47phox isoforms possesses four strictly conserved residues (Pro-363, Pro-366, Arg-368, Pro-369; human-p47phox numbering), and the C-tail region had an additional four identical amino acids (Ile-374, Arg-377, Cys-378, and Thr-382; human-p47phox numbering) (Figure 3D). Consistent with these identities, the essential binding roles of three residues (Pro-363, Pro-366, Arg-368, indicated by black boxes of Figure 3D) were previously demonstrated by mutational analyses [18]. The C-tail region is thought to enhance the interaction between the PRR of p47phox and the SH3 domain of p67phox [18], since simultaneous mutation of six of these residues (Leu-373, Ile-374, Arg-377, Thr-382, Lys-383, and Lys-385; human-p47phox numbering, indicated by black boxes of Figure 3D) significantly decreased the binding of the p47phox PRR to the p67phox SH3 domain [18,72]. Four of these residues (Ile-374, Arg-377, Thr-382, and Lys-383) are highly conserved among both p47phox and NOXO1 proteins, but two other residues (Leu-373 and Lys-385; human-p47phox numbering) were conserved only in p47phox orthologs (Figure 3D). This suggests that the binding affinities of p47phox and NOXO1 toward their respective activator protein SH3 domains may differ due to differences in these residues. Ser-379 in the C-tail region of human p47phox is a phosphorylation site [19]; the phosphorylation of which negatively regulates binding of the PRR and C-tail regions to the p67phox SH3 domain [72]. The threonine or serine residue is conserved at the position in all vertebrate p47phox and *G. gallus*, *D. rerio* and *O. latipes* NOXO1 (indicated by an arrow head in Figure 3D), indicating that NOXO1 proteins of these species retain this feature of p47phox which is subsequently lost in higher vertebrate NOXO1.

T. rubripes C17orf39 ortholog as a candidate NOXO1-like protein

The NOXO1 gene seems to have been lost from the *T. rubripes* genome, as shown in Figure 2A. On the other hand, orthologs of chromosome 17 open reading frame 39 (C17orf39) gene predicted by the ensembl transcript model [73] are present adjacent to the NOXO1 genes in the *D. rerio* and *T. nigroviridis* genomes (Figure 2A).

Amino acid sequences of these C17orf39 orthologs showed conserved PRR and polybasic regions (Figure. 4B, respectively, indicated by black boxes). In addition to the PRR and polybasic region, the *T. rubripes* C17orf39 ortholog has an extended region that encodes domains analogous to the C-terminal regions of NOXO1; these contain the bis-SH3 domain, PRR and C-tail region (Figure 4A). The sequence of *T. rubripes* C17orf39 possesses the 22 residues that are conserved in bis-SH3 domains of p47phox and NOXO1 (Figure 4C) and the eight key residues conserved in the PRR and C-tail regions (Figure 4D). A phosphorylation site corresponding to Ser-379 residue of human p47phox is also conserved in the *T. rubripes* C17orf39 (Figure 4D). Although the function of C17orf39 has not been established in any species, its NOXO1-like domain structure suggests that the C17orf39 ortholog of *T. rubripes* may function in place of the authentic NOXO1.

Identification of putative critical amino acids conserved among p67phox/NOXA1 orthologs

Alignment of the amino acid sequences of the 24 Nox-activator proteins (including amoebal and fungal isologs) identified 13 conserved amino acids in the TPR domains (Figure 5A). This includes two residues that have been identified as essential for binding human p67phox to Rac, as determined by the crystal structure of the complex [21] (Figure 5A, filled boxes). The AD was first identified by mutagenesis of p67phox [30]. Val-204 of human p67phox and the corresponding residue Val-205 of human NOXA1 are crucial for respective activation of Nox2 and Nox1 [30,41]. All orthologs of p67phox and NOXA1, and one fungal NOXR (*A. nidulans*-NOXR) conserve this valine residue (Figure 5C). The two fungal NOXR (*M. grisea* and *F. graminearum*) retained the hydrophobic amino acid leucine instead of valine at this position (Figure 5C). Similarly, the second AD-like region of *S. purpuratus* p67phox-like protein (Sp-p67phox-C) conserved an isoleucine resi-

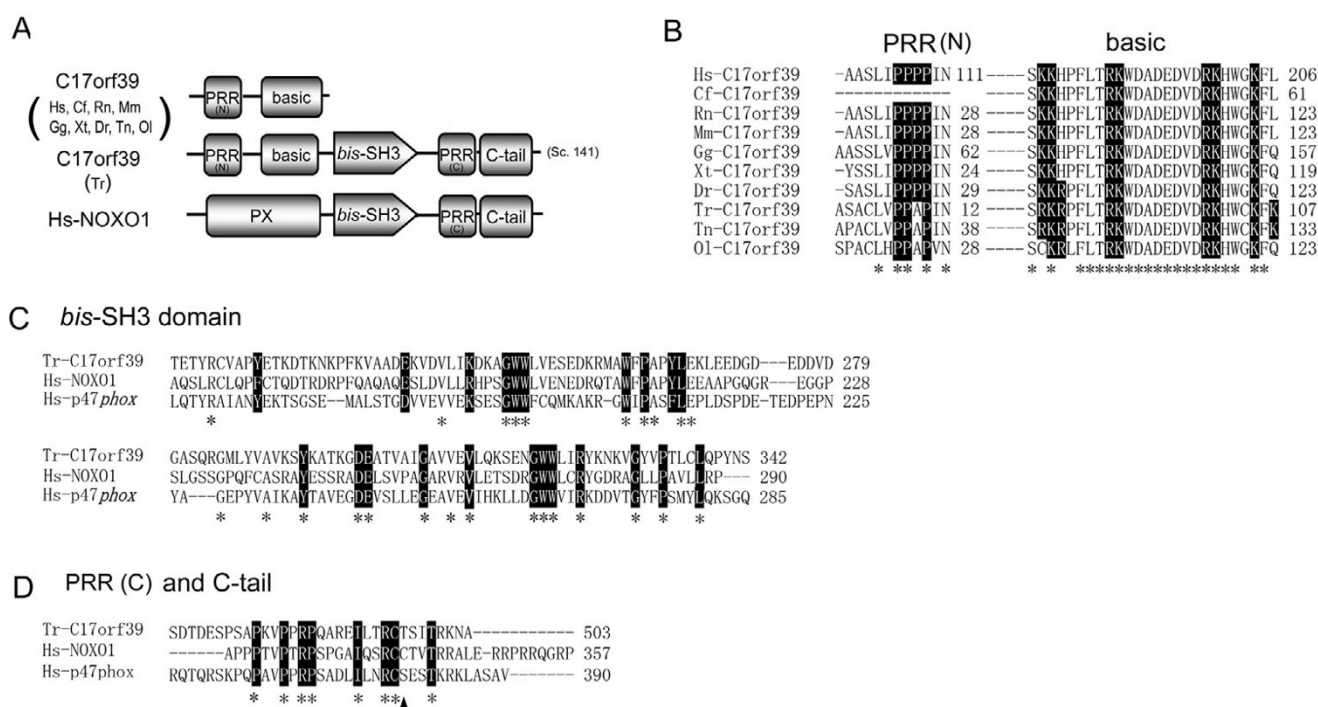


Figure 4

Hypothetical function of *T. rubripes* C17orf39 protein as a Nox organizer protein. (A) Schematic domain structures of C17orf39 and NOXO1. Names of orthologs that possess domains are shown to the left of structures. *T. rubripes* C17orf39 was located in scaffold 141 (Sc 141). Abbreviations of species names and most domains are shown in Figure 3; "basic" indicates a lysine/arginine-rich polybasic region. (B) Alignments of the PRR and basic regions of C17orf39. Letters in solid boxes indicate conserved the proline residues in the PRR and lysines and arginines in the basic region. (C) Comparison of bis-SH3 domain among *T. rubripes* C17orf39, human NOXO1, and human p47phox. (D) Comparison of PRR and C-tail regions among *T. rubripes* C17orf39, human NOXO1, and human p47phox. Letters in solid boxes indicate the evolutionally conserved residues that have been identified in Figure 3 (C, D). An arrow head indicates a residue corresponding to a phosphorylation site of human p47phox. Identical residues are shown by asterisks (B-D).

due at this position. In contrast, Dd-p67-like protein lacked a typical AD sequence (Figure 5C). The N-terminal region containing the TPR and AD can be considered as the "Nox-activating region", a module that cooperates with Rac to activate Nox enzymatic activity [11,29-31,74]. Taxonomy of the "Nox-activating region" (corresponding to residues 1–212 of human p67phox) reveals distinct subgroups corresponding to NOXA1 and to p67phox, and shows that the NOXR proteins of fungi also represent a distinct subgroup (Figure 5B). A fourth rather diverse subgroup is comprised of p67phox-like proteins that include those of *S. purpuratus* and *C. intestinalis* (numbers 19–21 in Figure 5B). Taken together with the taxonomy of p47phox (Figure 3B) and of Noxes [1], these data support an evolutionary model in which duplication of both the p67phox and p47phox genes occurred in vertebrates at a time corresponding to the appearance of Nox1. In contrast to fungal NOXR, Dd-p67-like protein appeared to branch from a root shared with sea urchin-p67phox-like genes (Figure 5B); however, the bootstrap value was only 35% raising uncertainty as to which subgroup the amoebal p67-like protein belongs. Nevertheless, amoebal p67-like protein is clearly distinct from the NOXR group.

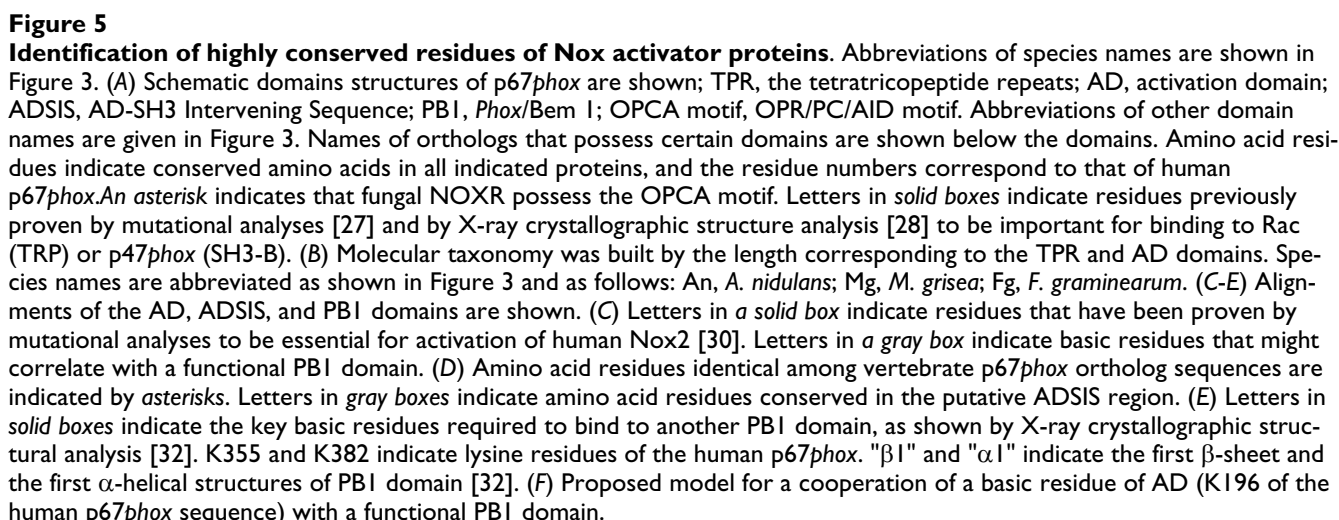
In contrast to the C-terminal SH3 domain (SH3-B) of activator proteins which is seen in all vertebrate activator proteins, the central SH3 domain (SH3-A) was seen only in vertebrate p67phox (Figure 5A). Comparison of sequences of all vertebrate p67phox orthologs identified 31 highly conserved amino acids in this SH3 domain (Figure 5A and Additional file 3). In addition, a region between AD and SH3-A (termed here the ADSIS region for "AD-SH3 Intervening Sequence") was strictly conserved among vertebrate p67phox isoforms, but was not seen in others, such as NOXA1 and NOXR. A consensus sequence was identified in this region (Figure 5D). A PRR-like region (residues 227–231 of human p67phox) was seen in ADSIS region, but it does not conserve in all vertebrate p67phox isoforms based on the deduced sequences (Figure 5D). The role of the putative region is currently unknown, but may involve a function related to SH3-A because it coincides with the presence of SH3-A. Despite the high conservation of SH3-A among vertebrate p67phox orthologs, the role of this domain is not defined [10,75].

The PB1 domain was present in all vertebrate p67phox, in vertebrate NOXA1 and in fungal NOXR and amoebal p67-like protein, but not in *C. intestinalis* and *S. purpuratus* p67phox-like proteins (Figure 5A). The crystal structure of the heterodimer between the PB1 domains of p67phox and p40phox domain demonstrated that Lys-355 and Lys-382 in human p67phox are essential to bind the PB1 domain of p40phox [32,76]. Residues corresponding to Lys-355 and Lys-382 of human p67phox are conserved not only in the vertebrate p67phox, but also in the NOXA1

orthologs of chicken (*G. gallus*) and fish (*D. rerio* and *O. latipes*) (Figure 5E). This suggests that in addition to its well-described role in activating Nox2, p40phox probably also participates in activating Nox1 in chicken and fish (Figure 5E). On the other hand, NOXA1 of *X. tropicalis* has lost both of these p40phox-interacting residues. Thus, these data support an evolutionary model in which p40phox originally participated in regulating both Nox1 and Nox2 in vertebrates, but after the appearance of mammals, Nox1 (but not Nox2) became independent of p40phox. The lack of a PB1 domain in both *C. intestinalis* and *S. purpuratus* p67phox-like proteins is consistent with the absence of p40phox orthologs in these species (Figure 1B).

The analysis of PB1 domain sequences in Nox activators also suggests an unexpected relationship between the AD and the PB1 domain. AD regions of all vertebrate p67phox contain a positively charged residue, e.g., Lys-196 of h-p67phox (indicated by gray boxes in Figure 5C). This basic residue is also completely conserved in NOXA1 orthologs of *G. gallus*, *D. rerio* and *O. latipes* (Figure 5C), which retain functional PB1 domains (Figure 5E), but not in the other NOXA1 and p67phox-like proteins (Figure 5C) (with the single exception of the C-terminal AD-like region of *S. purpuratus* p67phox-like protein). Interestingly, the fungal NOXR, which possess these key basic residues in their PB1 domains (Figure 5E), also conserves this basic residue in their AD regions (Figure 5C). Thus, the presence of a functional PB1 domain correlates strongly with the presence of a positively charged residue corresponding to position 196 in the human p67phox sequence, and may point to the functional cooperation of these two regions, e.g. in binding to p40phox or other functionally analogous proteins in fungi (Figure 5F). Alternatively, the positively charged residue in AD region might be related to the tail-to-tail interaction between vertebrate p47phox and p67phox (and NOXO1-NOXA1 of early vertebrates) because a sequence analysis of Nox organizers that is shown in Figure 3D suggests that the conserved residue also co-occurs with the phosphorylation site in the C-tail region of all vertebrate p47phox and early vertebrate NOXO1. Thus, co-appearance and co-disappearance of evolutionarily conserved residue provides a hint to find an unknown relationship between regulatory-subunits.

SH3-B was seen in all vertebrate p67phox and NOXA1 orthologs and in *S. purpuratus* p67phox-like protein (Figure 5A). Among these orthologs, seven conserved amino acid residues in SH3-B were identified (Figure 5A), and the residues include a tryptophan (Trp-494; human p67phox- numbering) that was previously shown to be crucial for interaction with the C-terminal PRR of Nox organizer proteins [40].



Identification of putative critical amino acids conserved among p40phox orthologs

Mammalian p40phox is considered to be an enhancer of Nox2 activation [33,35,77,78]. p40phox orthologs emerged in vertebrates (see Figure 1B) coincident with the appearance of the PB1 domain of p67phox-like proteins. Fungal NOXR and amoebal p67-like proteins also possess the PB1 domains (Figure 5E), but their genomes do not encode a distinct p40phox ortholog. The protein sequences of vertebrate p40phox orthologs are highly conserved; the canonical PX, SH3, and PB1 domains are all seen among vertebrate p40phox orthologs (Figure 6A). Alignment of ten vertebrate p40phox orthologs showed 48 conserved amino acid residues in the PX domain (see Additional file 3), including Phe-35, Arg-57, Tyr-59, Leu-65, Arg-105 (human p40phox numbering) that have been demonstrated to be essential for phospholipid-binding [23,24,79,80]. Phylogenetic analysis using the trimmed PX domain sequences (Figure 6B) does not reveal obvious sub-families. Recently a novel intramolecular interaction in p40phox between the PX and PB1 domains was reported [81]. The interface between the PX and PB1 domains consists of a hydrophobic pocket by Phe-35 of the PX domain and Val-257, Pro-265, Leu-273 and Phe-320 of the PB1 domain and ionic interactions between His-38, Arg-58, and Arg-60 of the PX domain and Glu-259 and Asp-269 of the PB1 domain (human p40phox numbering) [82]. The residue corresponding to the Leu-273 is much varied among vertebrate p40phox proteins (see Additional file 3), suggesting that the residue is less crucial than that previously proposed if the interaction is important in other vertebrates than human. The residue to His-38 is often replaced by tyrosine. The majority of other residues are conserved, but there are some exceptions: residues corresponding to Arg-60 in *X. tropicalis* p40phox and to Glu-259 and Asp-269 in rat p40phox (see Additional file 3).

To the best of our knowledge, a role for the SH3 domain of p40phox has not been defined [10]. Eighteen amino acids are conserved in the SH3 domain (see Additional file 3). Between the PX and SH3 domains, we identified a new conserved region of 15 amino acids that is rich in proline, arginine and lysine, and is therefore designated the "proline-basic (P-basic) domain" (Figure 6A). This putative region is not essential for PtdIns(3)P-binding by the p40phox PX domain [25,80]. An intramolecular interaction between the PX and PB1 domains seem not to be related to P-basic region according to the structure of full length p40phox [81,82]. The structural analysis demonstrates that P-basic region forms a separate motif and is situated closely to the PX domain [82], indicating that it might support the cell membrane-binding function of p40phox together with the PX-domain. Alternatively, the P-basic region might interact with the SH3 domain of

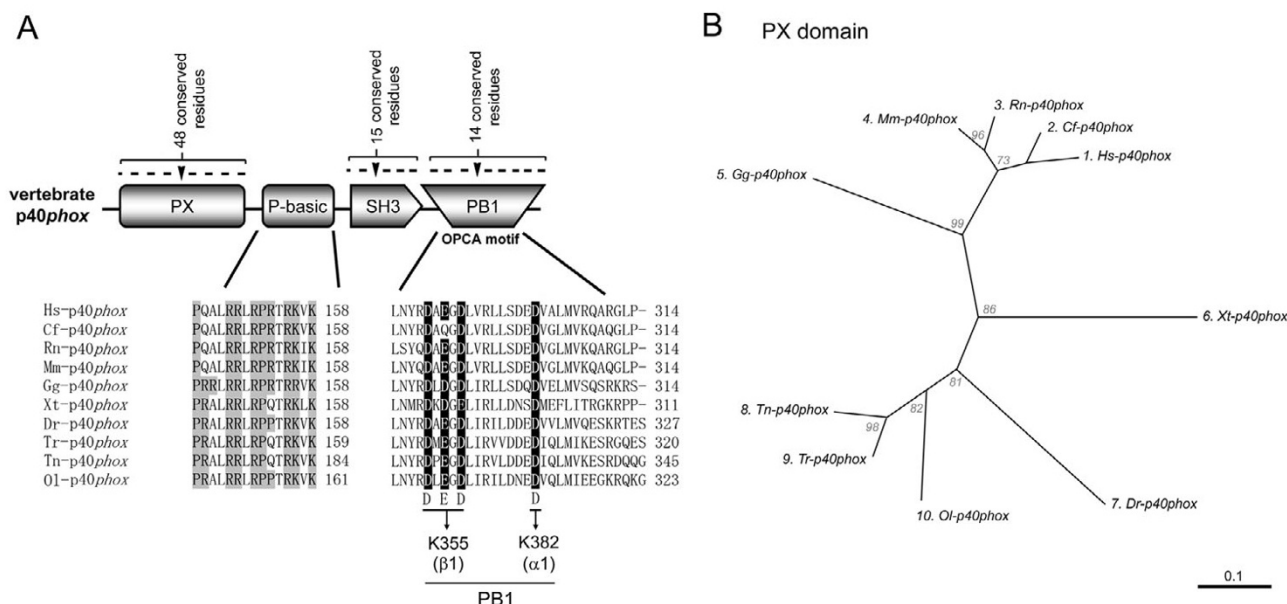
p40phox in a manner similar to the interaction between the AIR and the tandem SH3 of human p47phox. Interestingly, Thr-154 of human p40phox, located in the center of P-basic region, has been identified as a phosphorylation site [16], and this residue is entirely conserved among all vertebrate p40phox orthologs (Figure 6A).

The PB1 domain of human p40phox harbors acidic amino acids forming the "OPCA (QPR/PC/AID) motif". An OPCA motif contains four distinctive acidic residues that are essential for binding to conserved basic residues in the p67phox PB1 domain [32]. The four acidic residues, Asp-289, Glu-291, Asp-293 and Asp-302 (human p40phox numbering) are conserved in all orthologs with the exception of a glutamine rather than a glutamic acid in the position of *C. familiaris* p40phox that corresponds to Glu-291 of human p40phox (Figure 6A).

Identification of putative critical amino acids conserved among p22phox orthologs

Human p22phox contains two predicted transmembrane regions [83] and forms a stabilizing heterocomplex with Nox2 as well as Nox1, Nox3, and Nox4 [51-53]. The PRR of p22phox is the only motif in this protein whose role in Nox activation has been demonstrated [3,10]. The PRR is highly conserved among all p22phox orthologs (Figure 7A, the sequences are provided in Additional file 4), consistent with its role as a docking site for the bis-SH3 domain of organizer proteins. In addition, a total of 38 amino acids were identical in all p22phox sequences (Figure 7A), suggesting important additional functions, particularly for the N-terminal regions preceding TM1. Within these conserved residues, we identified a region rich in arginine and lysine (Figure 7B), and is therefore designated the polybasic region (indicated as "basic" in Figure 7A). A short sequence containing multiple basic amino acid residues has been proposed to contribute to recruitment of proteins to membrane sub-regions and to function in receptor-internalization via interaction with phospholipids [84-86]. According to our preliminary data, mutation in the polybasic region of p22phox did not affect extracellular ROS production from Nox2-transfected human embryonic kidney 293 cells (T. Kawahara and J. D. Lambeth, unpublished observations). This implies that the putative conserved region might function in a role unrelated to the enzymatic activity of Nox2, e.g. in phagocytosis or receptor-mediated endocytosis.

As shown in Figure 1, a p22phox-like protein was not identified in *S. purpuratus*, but because the genome sequence of this organism is incomplete and because a p47phox-like protein with a conserved bis-SH3 domain is present, we suggest that this organism probably possesses an undiscovered p22phox-like protein, or a protein that serves an analogous function. Our analysis did not provide con-

**Figure 6**

Identification of highly conserved residues of p40phox. (A) A schematic structure of p40phox domains is shown; proline-basic region, P-basic. Abbreviations of domain names are described in legends to other Figures. Letters in gray boxes indicate highly conserved proline and basic amino acid residues in the P-basic region. Letters in solid boxes indicate key acidic residues of OPCA motif that have been demonstrated by X-ray crystallographic structure [32]. K355 and K382 indicate two residues of human p67phox that interact with the OPCA motif. (B) Molecular taxonomy was generated by the alignment of amino acid sequences of PX domain. Species name abbreviations are given in Figure 3.

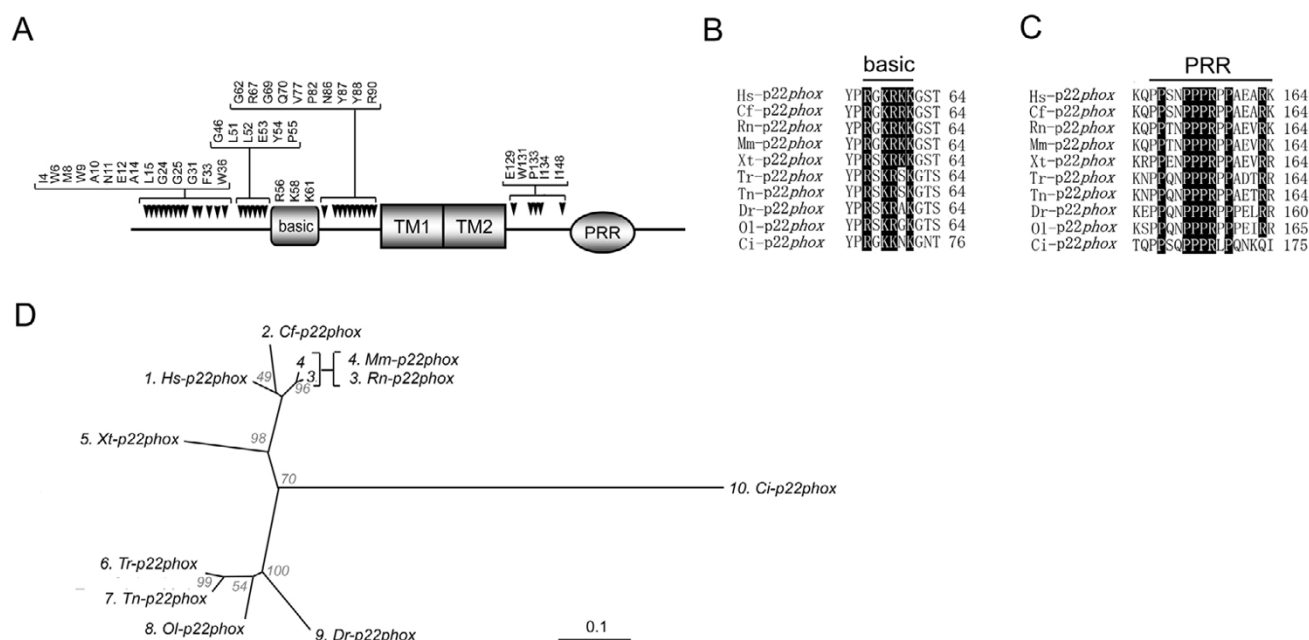
vincing evidence for a p22phox isolog in slime mold amoeba. Lardy *et al.* argued that this organism may possess a membrane-associated p22phox-like protein (GenBank™ accession number AY221170); predicted p22phox-like gene-null *D. discoideum* mutant is unable to produce spores, similar to *nox*-null mutants [56]. However according to our analysis, this protein is highly divergent in amino acid sequence compared with the consensus sequence obtained from all other organisms, showing disagreement in 24 out of the 38 conserved residues shown in Figure 7A. Thus, further investigation is needed to judge whether the gene emerged as a later adaptation or whether it represents a primitive p22phox gene.

Molecular evolution of domains of Nox organizers and activators in Deuterostomia

As described above, comparisons of the amino acid sequences of the Nox regulatory subunit orthologs revealed certain features of the evolution of characteristic domains. Some key residues and domains are summarized in Figure 8, which shows the proposed evolution of organizer and activator proteins. The p47phox orthologs of all vertebrates (top four left-hand panels in Figure 8) have identical domain structures, which include the PX domain, bis-SH3 domain, AIR, and PRR. On the other

hand, the domain composition of NOXO1 (right-hand panels in Figure 8) changed during the evolution from fish to tetrapods: the NOXO1 orthologs of tetrapods and fish all possessed the PX domain, bis-SH3 domain, and PRR. However, the predicted AIR (including key conserved residues, for details, see Figure 3C) is also present in *O. latipes* but is absent in *D. rerio* and tetrapods (right-hand panels in Figure 8). Despite the presence of the AIR in *O. latipes* NOXO1, it is clearly distinguished by sequence homology and synteny from a p47phox isolog, as shown in Figure 3B. Thus, initially, following gene duplication from p47phox, NOXO1 retained the AIR regulatory feature of p47phox, and this feature was subsequently lost.

Like vertebrate isologs of p47phox, the p47phox-like proteins of *C. intestinalis* and *S. purpuratus* possess the PX domain, bis-SH3 domain, and AIR. However, these isologs differ considerably in the C-terminal regions that, in higher forms, interact with p67phox isologs. The C-terminus of the *C. intestinalis* p47phox lacks the PRR, but motif scanning by PROSITE [87] demonstrates the p47phox-like protein contains, instead, additional SH3 domain [residues 340–399 of Ci-p47phox protein, GenBank™ No. [NM_001033828](#)]. The *S. purpuratus* p47phox-like protein

**Figure 7**

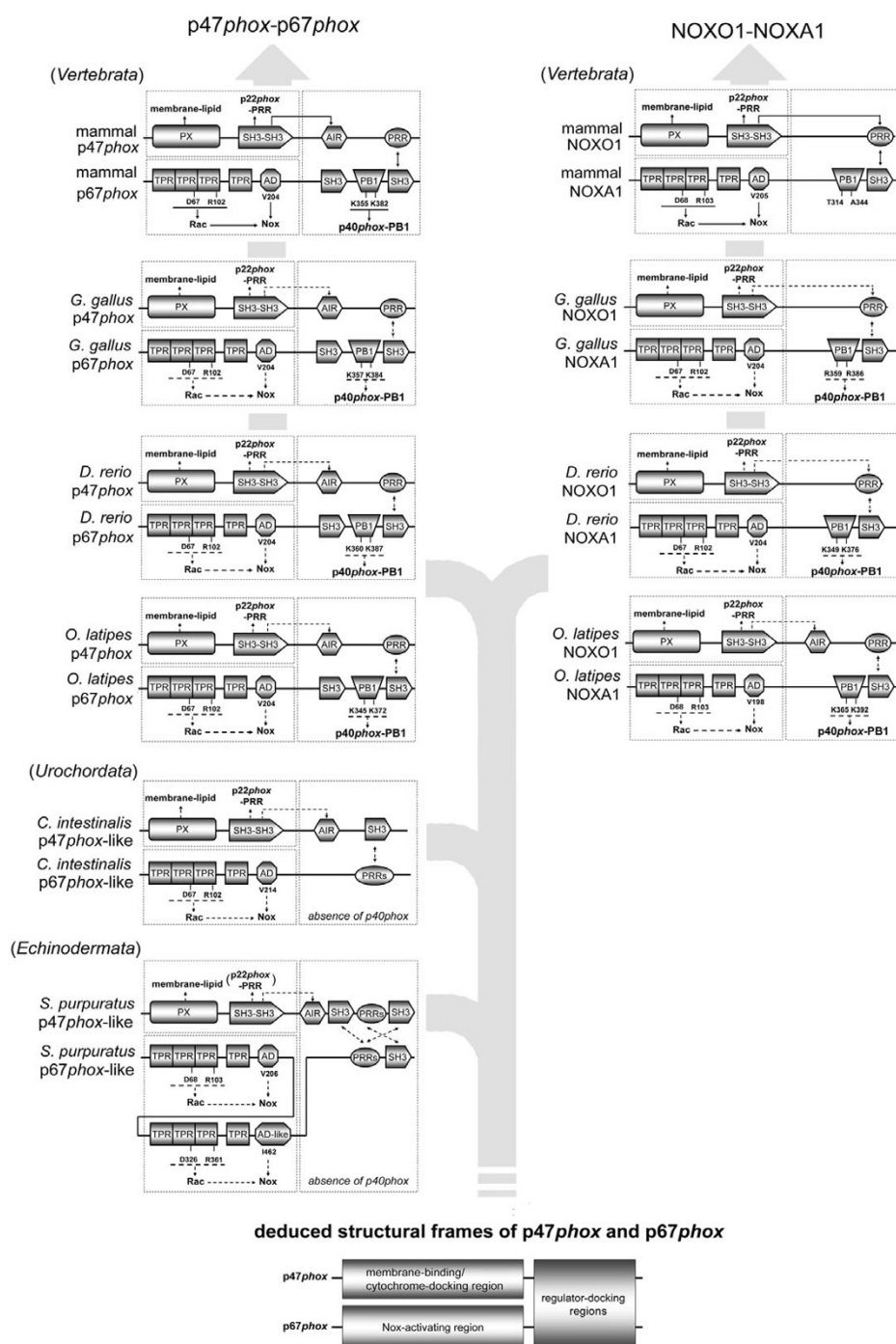
Identification of highly conserved residues of p22phox. (A) Schematic domain structures of p22phox are shown; TM1 and TM2 represent the predicted 1st and 2nd transmembrane region; PRR, proline-rich region; basic, a lysine- and arginine-rich conserved domain. Indicated above the schematic structure are putative key amino acid residues that are conserved in p22phox orthologs, and the residue numbers correspond to those of human p22phox. Detailed alignment of the polybasic region (B) and the PRR (C) are shown. Letters in solid boxes indicate the residues that have been proven by mutational analyses to be important for binding to p47phox and/or activation of human Nox2 [53, 75, 127, 128]. (D) Molecular taxonomy was generated based on the alignment of amino acid sequences of full-length p22phox. Species name abbreviations are given in Figure 3.

also lacked the C-terminal PRR, but instead had two additional SH3 domains (residues 367–430 and 561–618 of Sp-p47phox, GenBank™ No. XP_001183696) separated by two PRRs (residues 453–460 and 476–484 of Sp-p47phox) (bottom two left-hand panels in Figure 8).

All vertebrate orthologs of p67phox have the same domains seen in human p67phox (Figure 8, left panels). Importantly, all p67phox and NOXA1 proteins possess the AD region, which has been shown to be critical for activating superoxide generation in Nox2 and Nox1 [30,31]. The TPR motif is present in all species, and the two critical residues for Rac-binding, Asp-67 and Arg-102 of human p67phox [21], are conserved among all vertebrate isoforms of p67phox and NOXA1 (top four panels in Figure 8), supporting the idea that binding to Rac is a conserved feature of vertebrate p67phox and NOXA1. The N-terminal domain structures of *C. intestinalis* and *S. purpuratus* p67phox-like proteins are similar to those of vertebrate p67phox and NOXA1 (Figure 8). Both p67phox-like proteins possess a TPR domain retaining the two critical Rac-binding residues (Figure 8). In addition, *S. purpuratus* p67phox-like protein possesses a second set of TPR and

AD-like domains. Both TRP domains retain conserved essential Rac-binding residues, but the second AD-like domain was not well conserved as shown in Figure 5C. Thus, Rac binding is likely to be an evolutionarily conserved feature among all p67phox/NOXA1 proteins.

In mammals, binding between the PB1 domains of p67phox and p40phox has been shown to mediate heterodimer formation. The crystal structure of the heterodimer of the PB1 domains from p67phox and p40phox [32] demonstrated that Lys-355 and Lys-382 in human p67phox are essential to bind to the PB1 domain of p40phox, and mutation of Lys-355 of human p67phox abolished binding to p40phox [76]. These crucial residues are entirely conserved in the PB1 domains of all vertebrate p67phox proteins, which imply that they all bind to p40phox (top four left-hand panels in Figure 8). In contrast to vertebrate p67phox, mammalian NOXA1 does not retain these two key amino acids (a top right-hand panel of Figure 8), explaining the failure of NOXA1 to bind to p40phox [40]. However, the chicken *G. gallus*, and the fish *D. rerio* and *O. latipes* NOXA1 completely conserve these two residues, and an OPCA motif of p40phox (Figure 6),

**Figure 8**

Evolution of domains of p47phox /NOXO1 and p67phox /NOXA1 orthologs. Shown is a schematic representation of identified domains and known or predicted inter- and intra-molecular interactions among p47phox/NOXO1 and p67phox/NOXA1 orthologs in "mammals" (*H. sapiens*), *G. gallus*, *D. rerio*, *O. latipes*, *C. intestinalis*, and *S. purpuratus*. Solid lines indicate interactions that have been demonstrated experimentally. Broken lines indicate predicted or proposed interactions based on sequences analysis and correspondence to experimentally demonstrated interactions in other species. The inferior diagram indicates deduced generalized structural features of p47phox- and p67phox-like proteins. Abbreviations of domain names are given in legends to other Figures.

which is essential for binding to the p67phox PB1 domain are highly conserved. The conservations of key residues suggest that in these fish, binding to p40phox may be retained (from the second to fourth panels in right hand of Figure 8). Thus, the loss of function of p40phox can be traced in the evolution of NOXA1. Both *C. intestinalis* and *S. purpuratus* p67phox-like proteins lack a PB1 domain, consistent with the absence of p40phox orthologs.

The PX-domain of human p40phox specifically binds to PtdIns(3)P [23,24,79,80]. Therefore, p40phox accumulates during phagocytosis at early endosomes, which are rich in PtdIns(3)P [35,81,88]. *T. rubripes* p40phox is expressed in a wide range of tissues including the gastrointestinal tissues [89], while Nox2 expression is restricted to the blood and kidney. Although Nox1 expression in fish or the other non-mammal organisms is not clear, our analysis suggest that Nox1 of fish and birds might function, along with the p40phox orthologs, in endocytosis, e.g., in receptor-internalization. This novel relationship between Nox1 and p40phox may provide a clue to understanding the physiological role of Nox1 in vertebrates.

The vertebrate p67phox and NOXA1 orthologs all have the C-terminal SH3 domain that, in human, mediates binding to the PRR of p47phox and NOXO1, respectively. This supports the idea that all of the vertebrate forms of these proteins mediate protein interactions via tail-to-tail binding between the SH3 domain of the activator protein and the PRR of the organizer protein. We further suggest that even though the structures of the C-termini of p47phox- and p67phox-like proteins in *C. intestinalis* and in *S. purpuratus* differ from the vertebrate proteins, the same sort of tail-to-tail binding of these p67phox-like proteins to the corresponding p47phox-like proteins occurs as diagrammed at the bottom of Figure 8. For example, we suggest that the C-terminal putative PRRs of *C. intestinalis* p67phox-like proteins (residues 256–348 of Ci-p67phox protein, GenBank™ No. NM_001033827) binds to the C-terminal SH3 domain of the p47phox-like protein as indicated by a broken line in Figure 8, reversing the mode of interaction seen in the vertebrate proteins but accomplishing the same function. Similarly, SH3 domains and putative PRRs in the C-terminus of *S. purpuratus* p47phox-like protein may interact with the PRRs and the SH3 domains in the C-terminus of *S. purpuratus* p67phox, respectively. Thus, while domains in the C-termini of p47phox and p67phox orthologs are not conserved during evolution, the model shown in Figure 8 suggests that the "docking function" has been retained. The model suggests three functional modules within the organizer protein/activator protein complex: 1), a "membrane-binding/cytochrome docking region" on the organizer protein that is comprised of the phospholipids-binding PX domain and the tandem SH3 region that binds to the PRR of

p22phox; 2) a "Nox-activating region" that contains the Rac-binding TPR domain and the AD, which together cooperate to stimulate the activity of the Nox catalytic domain; and 3) a "regulator-docking region" which uses evolutionarily variable but paired docking modules that link the "membrane-binding/cytochrome docking region" of p47phox to the "Nox-activating region" of p67phox (bottom panel in Figure 8).

Occurrence and origin of Phox-like regulatory subunits during evolution of Metazoa and Choanoflagellata

In addition to regulatory subunit-dependent Nox proteins of the Deuterostomia, we assembled predicted regulatory subunits and the subunit-regulated Nox isolog sequences present in the genomes of following species: the sea anemone *Nematostella vectensis* (*N. vectensis*), the limpet *Lottia gigantea* (*L. gigantea*), the marine choanoflagellate *Monosiga brevicollis* (*M. brevicollis*) (see Additional files 5, 6, 7, 8). According to the recent phylogeny based on small ribosomal subunit gene and other molecular tools [90], the bilaterian animals consist of three clades: Deuterostomia, Lophotrochozoa, and Ecdysozoa. A gastropod snail, the limpet *L. gigantea*, belongs to the Lophotrochozoa, and p67phox, p47phox, p22phox-like genes are found in this genome (Figure 9A). The predicted motif structures by the Pfam program are shown in Figure 9B; characteristic motifs of p67phox (TPR domain and AD-like region) and p47phox (PX domain and AIR-like region) as well as the key residues (detailed alignments are shown in Additional file 8) are conserved. The genome of *L. gigantea* also possesses an ancestral Nox2 gene as a co-ortholog of vertebrate Nox1-3 (Figure 9A and see Additional file 6) like Nox2 isologs in primitive deuterostomes [1]. On the other hand, any distinct regulatory subunit genes were not found in seven species that belongs to Ecdysozoa including two nematodes (*Caenorhabditis elegans* and *Caenorhabditis briggsae*), four insects (*Drosophila melanogaster*, *Apis mellifera*, *Anopheles gambiae*, *Aedes aegypti*), and one non-insect arthropod (the waterflea *Daphnia pulex*) (Figure 9A).

Before the emergence of bilateral animals, cnidarians split from the main animal lineage and the genome retained many of the features present in last common ancestor of animals [90]. The starlet sea anemone *N. vectensis* that belongs to the Cnidaria, conserves p47phox-, p22phox-like genes, and two ancestral Nox2 genes, Nox2A and 2B (Figure 9A). The characteristic motifs of the putative regulatory subunits are predicted by the Pfam program and shown in Figure 9B (alignments are shown in Additional file 8). *N. vectensis* p47phox possesses PX and bis-SH3 domains. The isolog also possesses a putative polybasic region within the sequence corresponding to the AIR of the human p47phox gene. A polybasic region is a class III consensus motif for binding to a SH3 domain [91]; there-

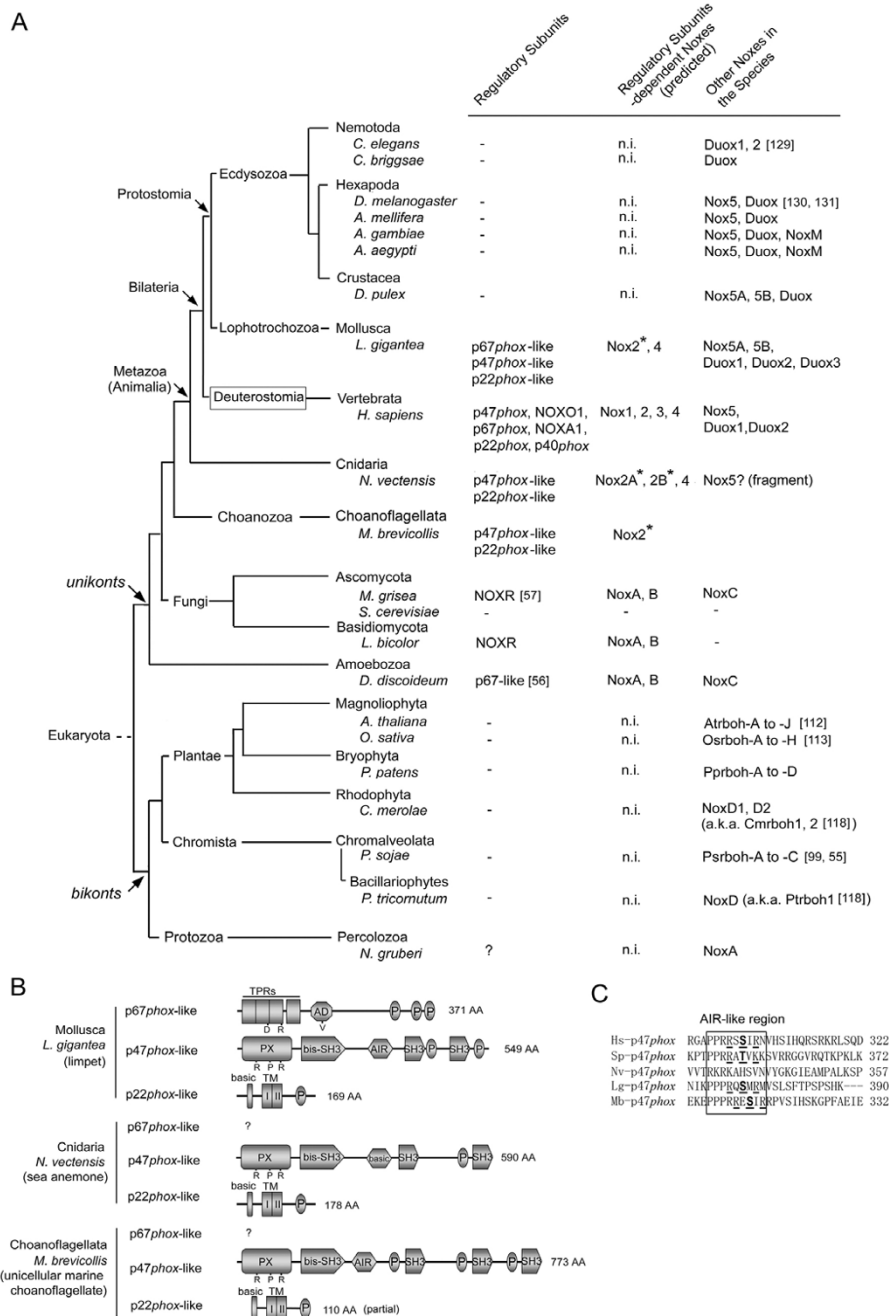


Figure 9
Relationship between the evolution of eukaryotes and the occurrence of *Phox* -related regulatory subunits. (A) A phylogeny of eukaryotes was created based largely on genetic information [90, 100]. Branch lengths are for illustrative purposes and are not proportional to phylogenetic distances of groups. Details of deuterostomes are shown in Figure 1. "n.i." indicates none identified or reported. "co-ortho. of v-Nox1-3" indicates that the gene is a co-ortholog ancestor of all members of the vertebrate Nox1-3 subgroup. The number in a parenthesis (e.g. [129, 130, 131]) indicates a reference number. (B) Predicted domains structures of newly identified putative regulatory subunit-like proteins are shown; P, proline-rich region. The sequences are provided in Additional file 7. Abbreviations of domain names are described in legends to other Figures. (C) Sequences of putative AIR-like region are shown. *Underlines* indicate residues that agree with a consensus sequence of protein kinase C substrate. *Bold letters* indicate predicted phosphorylation sites in AIR-like regions. Abbreviations: Hs, *H. sapiens*; Sp, *S. purpuratus*; Nv, *N. vectensis*; Lg, *L. gigantea*; Mb, *M. brevicollis*.

fore, this may implicate the polybasic region as a prototype of the AIR. Although the current version of the *N. vectensis* genome does not encode an obvious p67phox-like gene, the presence of multiple SH3 domains in *N. vectensis* p47phox points to an unidentified p67phox ortholog or an unknown PRR-containing binding partner. Although cnidarians have no specialized immune cells, the *N. vectensis* genome possesses orthologous genes to Toll/Toll-like receptors, complement C3-like molecules, and a redox-sensitive transcription factor nuclear factor- κ B that are orthologs of key components of innate immunity in vertebrates [92]. Further study needs to identify an original role of Phox-type NADPH oxidase in the primitive multicellular animals.

One clear implication of these genomic and detailed sequence analyses described above is that the hypothetical ancestor of primitive animals might conserve the Phox-type NADPH-oxidase system. In the middle of the nineteenth century, a striking structural resemblance between the collar cells, so-called "choanocytes", of primitive animal sponges and a group of protists the choanoflagellates has been noted ([93], reviewed in [94]). A line of recent phylogenetic analyses suggests that the metazoa including sponges, cnidarians, and other animals constitutes a monophyletic clade closely related to the Choanoflagellata [90,95,96]; therefore, the choanoflagellates are considered to be the organisms that are close in evolution to the all modern animals. The unicellular marine choanoflagellate *M. brevicollis* has been reported as a closer relatives to the hypothetical ancestor of all multicellular animals based on the comparative analyses of animal-specific genes [97,98]. Interestingly, the genome of *M. brevicollis* possesses p47phox- and p22phox-like genes (Figures 9A,B), but a BLAST searching in the current *M. brevicollis* genomic database failed to identify a distinct p67phox ortholog. Furthermore, Nox2-like gene is present in the *M. brevicollis* genome sequence (Figure 9A). Interestingly, a taxonomy of Nox domains demonstrates that *M. brevicollis* Nox2-like gene diverges from the monophyletic clade of all metazoan Nox2 genes containing other ancestral Nox2 and all vertebrate Nox1-3, but it differs from Nox4 and fungal NoxA/NoxB subfamilies (see Additional file 6). The *M. brevicollis* p47phox-like protein possesses AIR-like region that conserves a serine residue consistent with a consensus motif of a putative substrate of protein kinase C (Figure 9C); therefore, this implies that ROS-generating activity of the most primitive Nox2-like protein is also strictly regulated by phosphorylation as seen in mammalian phagocyte NADPH-oxidase. The physiological function of the primordial Nox2 enzyme in *M. brevicollis* is unknown, but the early appearance of the Phox-like Nox system in the unicellular flagellate suggests the presence of a fundamental and essential role of the regulatory subunit-dependent Nox in a unicellular living.

Figure 9A also summarizes the occurrence of Nox4, Nox5, and Duox isologs in genomes of the metazoa and the choanoflagellate, adding to information published previously [1]. Duox orthologs are present in all analyzed genomes of the Bilateria (Figure 9A). Nox5 genes occur broadly in the genomes of Metazoa, but not in Nematoda. Nox4 genes appear after the emergence of chordates, but is not seen in the echinoderm *S. purpuratus* [1] (Figure 1A). Interestingly, gene search that covers a wide range demonstrates that Nox4-like genes are also present in a lophotrochozoan (*L. gigantea*), a cnidarian (*N. vectensis*), but not in the ecdysozoans (see Figure 9A, and the taxonomy of Nox is shown in Additional file 6). The choanoflagellate *M. brevicollis* do not possess Nox4, Nox5, and Duox orthologs, at least based on the current genomic database. In the kingdom Plantae, the moss *Physcomitrella patens* (*P. patens*) possess double EF-hand-containing Noxes like the higher land plants *Arabidopsis thaliana* (*A. thaliana*) and *Oryza sativa* (*O. sativa*) (Figure 9A). Despite a kingdom distinct from plants, the oomycete *Phytophthora sojae* (*P. sojae*) also possesses single or double EF-hand(s)-containing Noxes [55,99], at least three genes (Figure 9A). Here we referred to them as *P. patens* and *P. sojae* rboh (respiratory burst oxidase homolog) to represent the remarkable resemblance to the higher plant Noxes, so-called rboh, in the respects of taxonomy of the Nox domains (Additional file 6). In the kingdoms Plantae and Chomista, a distinct regulatory subunit was not seen. However, higher plant genomes possess a novel protein family conserved domain structures similar to p67phox as described below. *Naegleria gruberi* (*N. gruberi*) is a widespread soil and freshwater amoeboid flagellate that belongs to the phylum Percolozoa, the kingdom Protozoa [100]. Although *N. gruberi* is a free-living organism, it is not related to species of the phylum Amoebozoa with sequenced genomes, such as *D. discoideum* and *Entamoeba histolytica*, but it is close to pathogenic relatives (e.g. *Naegleria fowleri*) that can cause amoebic meningitis. We identified single Nox gene of *N. gruberi* from the genome (Figure 9A), together with five putative *Fre* ortholog genes that have similar domain structures to that of Nox domain (see Additional file 9). According to a phylogenetic taxonomy of Nox domain, the *N. gruberi* Nox is close to *D. discoideum* NoxA/NoxB proteins (see Additional file 6); therefore, it is referred to as *N. gruberi* NoxA (Figure 9A). Nox regulatory subunit gene has not been found in the current version of genomic database, but it might contain a prototype of regulatory subunits according to the Nox taxonomy.

SH3PXD2 as a p47phox family member

As shown in Figure 9, a choanoflagellate and primitive animals possess p47phox-like genes as well as ancestral Nox2 and PRR-containing p22phox genes. On the other hand, further BLASTP search in the animal genomes using the *M. brevicollis* p47phox-like protein sequence suggested

the presence of a protein family proteins related to the primitive *p47phox* in higher animals, encoding the "SH3 and PX domains 2" protein (SH3PXD2) (sequences and alignments are provided in Additional files 10 and 11, respectively). The encoded protein is composed of an N-

terminal PX domain and a central *bis*-SH3 domain, plus two or three additional SH3 domains in the C-terminal half of the molecule (see the motif structure in Figure 10A). The SH3PXD2 gene is present in a urochordate, but was not seen in an echinoderm and other primitive meta-

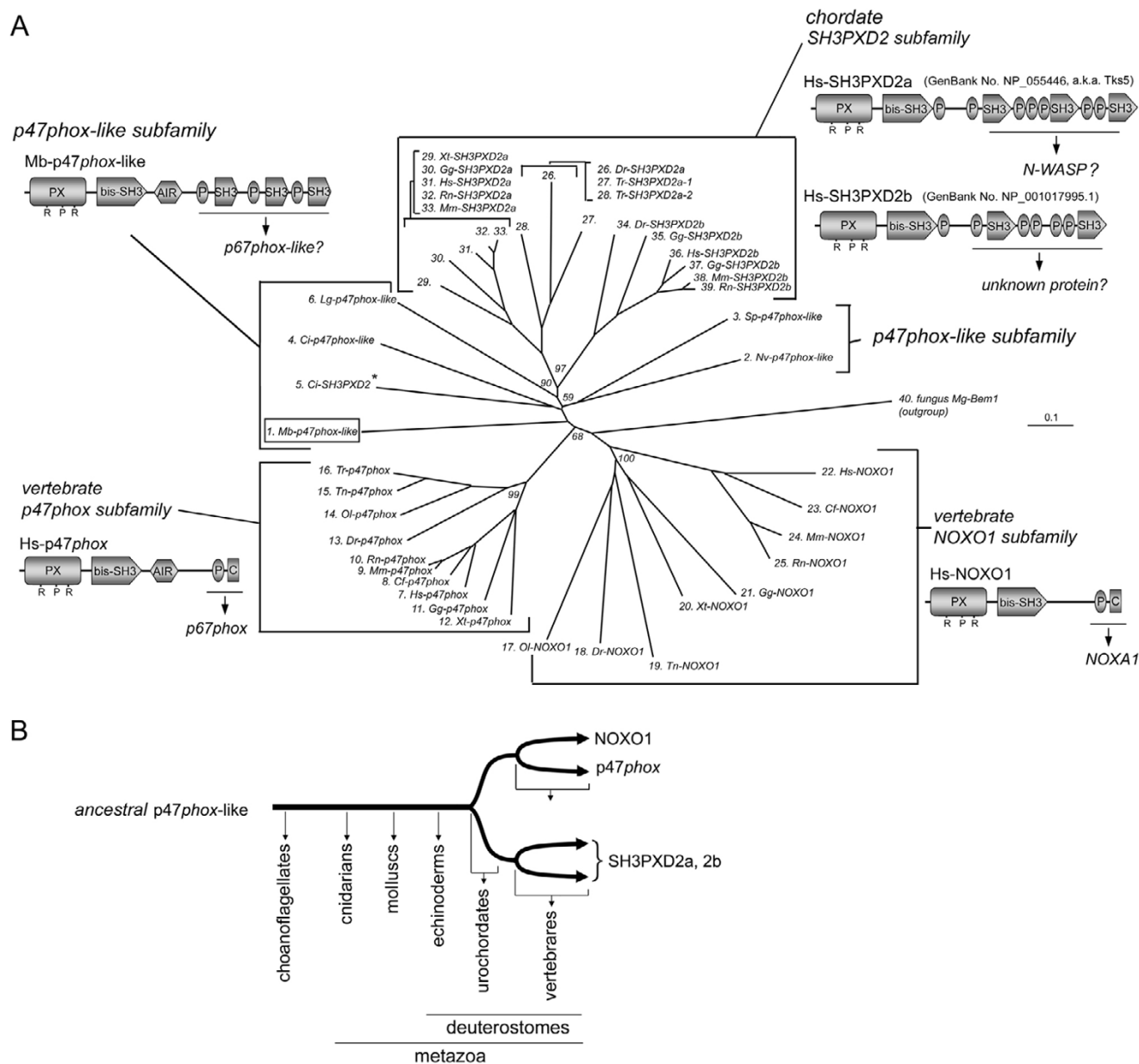


Figure 10

Evolution of a vast family of *p47phox* proteins. (A) Molecular taxonomy of the *p47phox* and related proteins is shown. The sequences of SH3PXD2 orthologs are provided in Additional file 10. Predicted domains structures of *H. sapiens* (*p47phox*, NOXO1, SH3PXD2a, 2b) and *M. brevicollis* (*p47phox*-like) proteins are shown near to each subfamily: P, proline-rich region; C, C-tail region. An asterisk indicates the appearance of a *C. intestinalis* SH3PXD2 branch from a common root of the *p47phox*-like protein. Abbreviations of species names and other domains are given in other Figures. (B) A schematic drawing to show a hypothetical evolution of an expanded *p47phox*-like gene family. Branches are for illustrative purposes and are not proportional to rates of divergence.

zoan species (Figures 10A and 10B). In vertebrates, two isozymes SH3PXD2a and 2b were found (Figure 10A). Interestingly the *M. brevicollis* p47phox-like gene and other primitive animal p47phox-like genes are related more closely to the SH3PXD2 gene subfamily than to the vertebrate p47phox and NOXO1 subfamilies (Figure 10A). The physiological function of SH3PXD2a is suggested by only a few papers; human SH3PXD2a protein (a.k.a. Tks5) is involved in the protease-driven invasion of cancer cells and the neurotoxicity induced by amyloid-beta [101,102]. A protein binding to SH3PXD2a is also little known, but recently N-WASP (Wiskott-Aldrich Syndrome protein) was proposed as a candidate that binds to the third SH3 domain of the human SH3PXD2a [101]. The physiological role as well as the binding partner of SH3PXD2b is unknown. However, the clear phylogenetic relationship of the SH3PXD2 gene family to the larger family of Nox-regulatory organizing proteins suggests that the genes might be involved in regulating Nox activity. Alternatively, SH3PXD2 proteins may have diverged to perform unrelated functions (e.g., to regulate other enzymes or signaling pathways). Thus, the *M. brevicollis* p47phox-like protein evolved from a hypothetical p47phox ancestor, and the SH3PXD2 genes diverged from the lineage of the p47phox-like gene following the emergence of chordates but prior to NOXO1 (Figure 10B). The vertebrate p47phox and NOXO1 were somehow able to eliminate the additional SH3 domains in the "regulator-docking region", as proposed in Figure 8.

What is the regulatory subunit binding partner for fungal NOXR?

Fungi (*A. nidulans*, *M. grisea*, and *F. graminearum*) belong to Ascomycota [55] and possess the NOXR gene as a p67phox ortholog (Figure 9A). NOXR encodes a PB1 domain homologous to that in p67phox, but these organisms do not possess a p40phox ortholog (see Figure 1A). As shown in Figure 5E, two important basic residues in the PB1 domains are conserved in all NOXR proteins. The conservation of these residues implies the existence of an unknown NOXR binding partner(s) containing a PB1 domain with an OPCA motif (Figure 11A). A BLAST search of a database of computationally predicted amino acid sequences [103] from the genomes of *A. nidulans*, *M. grisea*, and *F. graminearum* was performed using as a query sequence the PB1 domain of human p40phox (residues 237–339; GenBank™ No. [NP_000622](#)). In addition to NOXR, three sequences harboring PB1 domains were identified in the fungi. These were: "SH3-SH3-PX-SRR-PB1", "CDC24-RhoGEF-PB1" and "(CBS)n-PB1" proteins (Figure 11B and see Additional file 12), so-named for their domain structures in sequence from N- to C-termini. SRR indicates a serine-rich region. The three predicted proteins were encoded in all three fungal genomes, and all

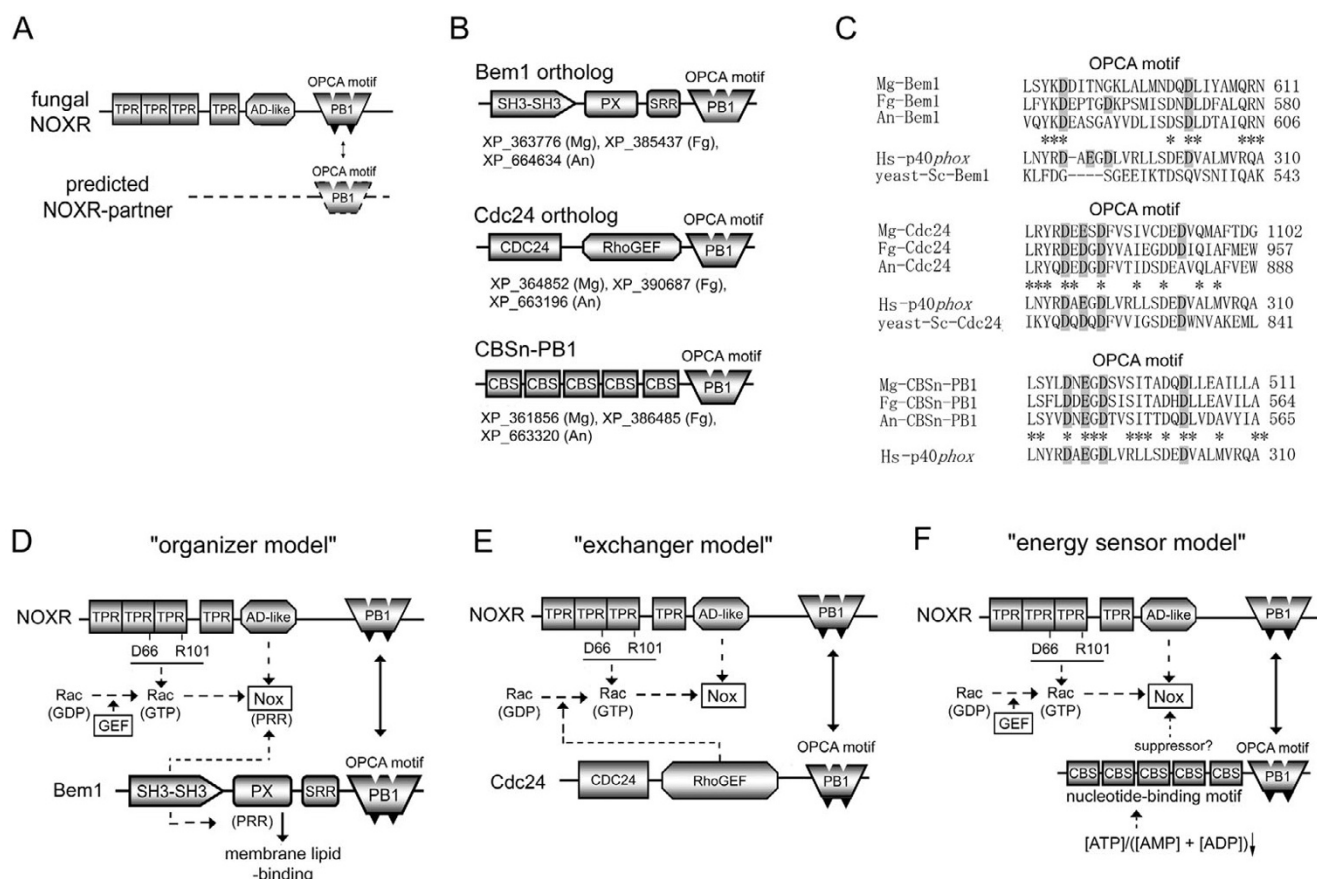
three possessed acidic OPCA motifs in their PB1 domains (Figure 11C).

The "SH3-SH3-PX-SRR-PB1" protein contains a similar domain structure to yeast *S. cerevisiae* Bem1p (Bud Emergence 1, GenBank™ accession number [NP_009759](#)), but differs in that the PB1 domain of *S. cerevisiae* Bem1p does not contain the OPCA motif [76]. A speculative model for the regulation of fungal Nox by the Bem1 ortholog is shown in Figure 11D, wherein Bem1 ortholog is proposed to play a role as an organizer protein, analogous to the roles of p47phox and p40phox in mammals. According to this model, PB1-PB1 interaction mediates the linkage between the "regulator-docking regions" suggested in Figure 8. PX-domain of the *S. cerevisiae* Bem1 facilitates a protein to anchor to a membrane lipid layer [104]. Fungi do not have a PRR-containing p22phox ortholog (see Figure 1A), but NoxA and NoxB in the three fungi has a distinctive highly conserved a class II PRR motif [91] (e.g. 423–428 residues of Mg-NoxA) and class III (e.g. 447–452 residues of Mg-NoxB) near the predicted NADPH-binding sub-regions [1]. Thus the hypothetical functions of the bis-SH3 and PX domains indicate the analogous function as a "membrane-binding/cytochrome docking region" of Nox organizer. Because *S. cerevisiae* Bem1 is involved in establishment of cell polarity [105], the proposed interaction between fungal NOXR and Bem1 orthologs may point to a function of Nox-derived ROS in cell polarity.

A second model is suggested by the "CDC24-RhoGEF-PB1" protein, which is an ortholog of *S. cerevisiae* Cdc24p (GenBank™ accession number [AAA82871](#)). According to this "exchanger model", the PB1-mediated heterodimer formation localizes the RhoGEF domain which then catalyzes the conversion of Rac-GDP to the active Rac-GTP (Figure 11E). The model is supported by a recent report showed that fungal NOXR binds to the fungal Rac homologue in the GTP form [57].

An "energy sensor" function for NOXR is shown in Figure 11F, and is based on the occurrence of the CBS (cystathione beta-synthase) domain in the putative binding partner. The CBS domain senses cellular energy status by binding adenosine nucleotides; it occurs in a range of proteins including AMP-activated protein kinase, which is activated by AMP and inhibited by ATP via the CBS domain [106,107]. Such a function might be related to the requirement of Nox enzyme for reduced pyridine nucleotide, thus affecting cellular energetics.

Fungal Nox and NOXR isologs are seen not only in fungi belonging to Ascomycota, but also some basidiomycetes [55]. To explore the likelihood of the fungal Nox activation mechanisms suggested in Figure 11, the occurrence of

**Figure 11**

Predicted binding partners of fungal NOXR and activation models of fungal Nox. (A) A hypothetical domain structure of a predicted NOXR partner is shown. (B) Domain structures of fungal proteins with PB1 domains meeting criteria for possible NOXR-partners are shown, along with their GenBank™ accession numbers. Abbreviations: An, *A. nidulans*; Mg, *M. grisea*; Fg, *F. graminearum*. (C) Aligned OPCA motifs of candidate NOXR partners are compared with the corresponding region in p40phox and the yeast isolog (the first and second alignments). Amino acid residues conserved among fungal ortholog sequences are indicated by asterisks. (D) A proposed model for activation of fungal Nox by NOXR and Bem1 protein. (E) A proposed model for activation of fungal Nox by NOXR and Cdc24. (F) A proposed model for activation of fungal Nox by CBSn-PB1 proteins. RhoGEF, Rho guanine nucleotide exchange factor; CBS, cystathionine beta-synthase; Abbreviations of other domain names are described in legends to other Figures.

NOXR, Bem1, Cdc24, and CBSn-PB1 isologs were compared in 13 fungal genomes spanning the four fungal divisions: Ascomycota, Basidiomycota, Zygomycota, and Chytridiomycota (sequences are shown in Additional file 12). Bem1, Cdc24, and CBSn-PB1 genes are present in most of the fungal genomes, with the exception of Cdc24 in *Batrachochytrium dendrobatidis* (*B. dendrobatidis*), and these proteins all conserved the PB1 domains with some exceptions; *Y. lipolytica* NOXR and *Coprinopsis cinerea* Cdc24 isologs lack the PB1 domain (Figure 12). In addition to the absence of Nox and NOXR isolog genes in *S. cerevisiae*, the Nox and NOXR isologs are absent in the genomes of *Candida albicans*, *Yarrowia lipolytica* (*Y. lipolytica*) and *Schizosaccharomyces pombe* (*S. pombe*) [55]. A Nox

gene is absent in the genomes of *Y. lipolytica* and *Phycomyces blakesleeanus* (*P. blakesleeanus*) despite the presence of NOXR isologs [55]. The *P. blakesleeanus* genome possesses two Bem1 and CBS-PB1 genes and all conserved the PB1 domains. We investigated the conservation of two basic residues of the PB1 domain corresponding to the Lys-355 and -382 residues of human p67phox and the four acidic residues in OPCA motif corresponding to the Asp-289, Glu-291, and Asp-293, -302 residues of human p40phox. Comparison of the PB1 domains in these fungal proteins by alignment and analyses by the Pfam program demonstrate that the two basic and four acidic residues are conserved in the fungal NOXR, Bem1, Cdc24, and CBSn-PB1 with a few exceptions; the OPCA motifs in Bem1 isologs

	NoxA	NoxB	NOXR	Bem1	Cdc24	CBSn-PB1
			(basic)(OPCA)	(basic)(OPCA)	(basic)(OPCA)	(basic)(OPCA)
Ascomycota						
<i>Sordariomycetes</i>						
<i>M. grisea</i>	✓	✓	✓ (RK)(DDGD)	✓ (KR)(DTGD)	✓ (KK)(DEDD)	✓ (KK)(DEDD)
<i>F. graminearum</i>	✓	✓	✓ (RK)(DDDD)	✓ (KR)(DTDD)	✓ (RK)(DDDD)	✓ (KK)(DEDD)
<i>Eurotiomycetes</i>						
<i>A. nidulans</i>	✓		✓ (RK)(D- DD)	✓ (KR)(DSAD)	✓ (KK)(DDDA)	✓ (KK)(DEDD)
<i>Saccharomycotina</i>						
<i>S. cerevisiae</i>				✓ (KR)(G- SQ)	✓ (RK)(DDDD)	
<i>C. albicans</i>				✓ (KR)(QPSN)	✓ (LQ)(DEDD)	✓ (KK)(DEDD)
<i>Y. lipolytica</i>			✓ (- -)(- - - -)	✓ (KR)(ADSD)	✓ (RK)(DDDD)	✓ (KK)(DEDD)
<i>Taphrinomycotina</i>						
<i>S. pombe</i>				✓ (KK)(DAKD)	✓ (RK)(DDDD)	✓ (QR)(DEDD)
Basidiomycota						
<i>Agaricomycotina</i>						
<i>C. cinerea</i>	✓	✓	✓ (RK)(DDGD)	✓ (KR)(DTTE)	✓ (- -)(- - - -)	✓ (KK)(DDDD)
<i>L. bicolor</i>	✓	✓	✓ (RK)(DDGD)	✓ (KR)(DTTE)	✓ (KK)(DDDD)	✓ (KK)(DDDD)
<i>P. placenta</i>	✓	✓	✓ (RK)(DDGD)	✓ (KR)(DTQD)	✓ (KK)(DDDD)	✓ (KK)(DDDD)
<i>Pucciniomycotina</i>						
<i>P. graminis</i>	✓	✓	✓ (RE)(GESD)	✓ (KR)(DSVP)	✓ (KK)(DEDD)	✓ (KK)(DEDD)
Zygomycota						
<i>P. blakesleeana</i>			✓ (RK)(DEED)	✓ (KR)(DTED) ✓ (KR)(DQED)	✓ (KK)(DDDD)	✓ (KK)(DEDD) ✓ (KK)(DEDD)
Chytridiomycota						
<i>B. dendrobatidis</i>	✓	✓	✓ (KK)(DEFD)	✓ (KR)(DCQD)	?	✓ (RA)(DEDD)

Figure 12

Conservations of NOXR, Bem1, Cdc24, and CBSn-PB1 genes and characteristic residues of PB1 domains in fungal genomes. (A) "basic" and "OPCA" in parenthesis indicate key residues that are required to form a dimer between two PB1 domains; two "basic" residues and four in "OPCA" motif residues correspond to Lys-355 and Lys-382 of human p67phox and Asp-289, Glu-291, Asp-293 and Asp-302 of human p40phox, respectively. Divisions of Fungi are shown in *bold*.

of *S. cerevisiae* and *C. albicans* has been lost (Figure 12 and the sequences and detailed alignments are shown in Additional file 13 and 14, respectively). The high conservation of the motif strongly implies that these PB1 domains mediate functionally important oligomerizations. Future functional studies will clarify the roles of these proteins in fungal Nox functions.

Predicted binding partners for D. discoideum p67-like protein

Like fungal NOXRs, the PB1 domain of *D. discoideum* p67-like protein also possesses two basic residues, Lys-313 and Lys-340 residues, as shown in Figure 5E. A BLAST search demonstrates that the *D. discoideum* genome possesses five predicted PB1 domain-containing proteins (referred to here as "Dd-PB1-A to -E"). All except Dd-PB1-D conserve the OPCA motif (Figure 13, and see details in Additional file 15). Dd-PB1-A protein (GenBank No. [XP_639165](#)) is partially identical to *D. discoideum* MEK kinase α (GenBank No. [AAC97114](#)) [108]. It is possible that the serine/threonine kinase domain might phosphorylate the amoebal Noxes or the p67-like protein to stimulate ROS generation, analogous to other regulatory

phosphorylations of Nox and their regulatory subunits [20,109-111]. Another interesting point of the Dd-PB1-A protein is to contain the F-box and a WD40 repeat regions that function as a ubiquitin ligase component E3. Further investigations to identify a binding partner will lead to understand functional relationships between Nox and the Dd-PB1 proteins.

Do p67phox-like genes exist in plants?

The land plants, *A. thaliana* and *O. sativa* possess the multiple EF-hand-containing Nox proteins, At-rboh-A to -J [1,112,113] and Os-rboh-A [114] to -H, respectively. However, there is no report of *Phox*-related regulatory subunit homologs in plant genomes. Unexpectedly BLAST search using PB1 domain sequences demonstrates that *A. thaliana* possesses a unique family of genes encoding four proteins that possess both TPR and PB1 domains (GenBank accession No. [NP_194935](#), [NP_180101](#), [NP_197536](#), [NP_564794](#)). The *O. sativa* genome also possesses a similar gene (GenBank accession No. [BAD31284](#)). Herein, we refer to this family as "hypothetical p67phox-like (hypo-p67-L)". The expressions of at

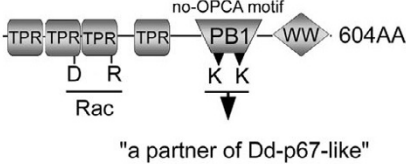
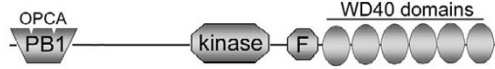


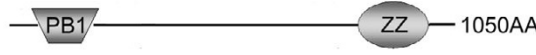

	name	GenBank™ No.	domain structure	(basic)(OPCA)
Amoebozoa				
Mycetozoa				
	<i>D. discoideum</i>	Dd-p67-like		(KK)(QDEQ)
		Dd-PB1-A (XP_639165)		(KD)(DEDD)
		Dd-PB1-B (XP_646538)		(KF)(DDDD)
		Dd-PB1-C (XP_644298)		(KI)(DEED)
		Dd-PB1-D (XP_638776)		(NK)(NDEH)
		Dd-PB1-E (XP_635325)		(KL)(DDDD)

Figure 13
Predicted candidate binding partners of *D. discoideum* p67-like protein. Predicted domain structures of Dd-p67-like protein and candidates of binding partners are shown. "basic" and "OPCA" in parenthesis indicate six characteristic residues of PB1 domain as shown in Figure 12. Abbreviations: kinase, protein serine/threonine kinase domain; F, F-box; WD40, WD40 repeat; WW, WW domain; ZZ, ZZ type zinc-finger.

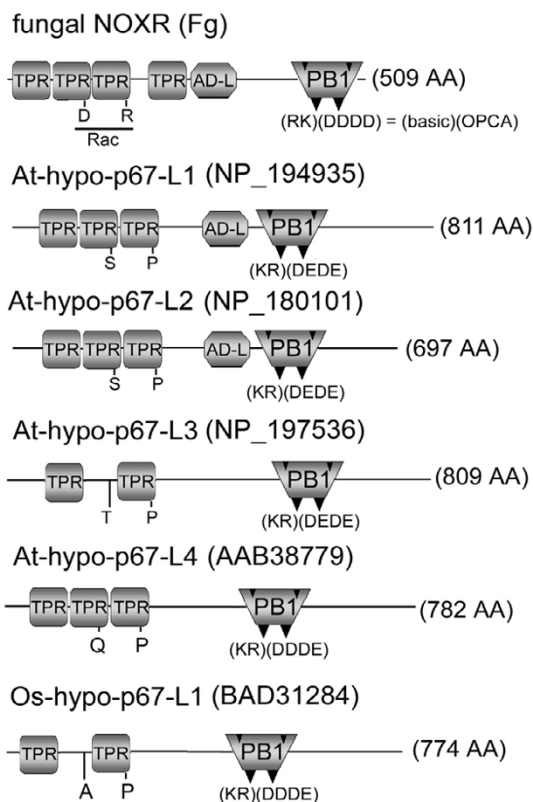
least the two identified genes, At-hypo-p67-L1 and -L4, were confirmed according to the EST database [115].

Domain structures of the *A. thaliana* and *O. sativa* hypo-p67-L proteins are shown in Figure 14A (sequences are provided in Additional file 16). The six residues characteristic of the PB1 domains are conserved in all sequences (Figure 14A, and see the alignment in Additional file 17). This predicts that the hypo-p67-L proteins will form a heteromeric or homomeric complex via PB1-PB1-interaction. In addition to the hypo-p67-L gene family, the *A. thaliana* genome possessed at least 27 isolated genes encoding the PB1 domain [e.g. protein kinase family protein (GenBank No. [NP_200569](#)), CBSn-PB1 ortholog (GenBank No. [NP_181191](#)), see Additional file 18]. Alignment of putative AD-like domains of the *A. thaliana* proteins with human p67phox and NOXA1 show that the *A. thaliana* hypo-p67-L1 and -L2 proteins conserve the functionally critical valine residue (Figure 14B). Moreover, a lysine residue corresponding to Lys-196 of human p67phox that is correlated to the appearance of functional PB1 domain is also conserved in two of them (Figure 14B). On the other hand, the sequences do not conserve residues in the TPR domain known to be essential for binding to Rac [10]. Biological functions for this putative family are unknown and further investigations are needed.

Conclusion

In summary, we reported here an exhaustive analysis of the *Phox*-related regulatory subunits for Nox enzymes from a total of 32 species in the Eukaryota including 10 vertebrates. The present study, viewed in the context of the evolution of Noxes [1], reveals that regulatory subunits co-evolved with their respective Noxes; Nox2 with p67phox/p47phox; Nox1 with NOXO1/NOXA1; ancestral Nox2 as co-ortholog of vertebrate Nox1-3 with p67phox/p47phox-like proteins; fungal and amoebal NoxA/B with NOXR (and possibly new components). By comparing amino acid sequences of the regulatory subunits, putative key amino acid residues and conserved region previously undefined were identified, e.g. ADSIS. These conserved residues are likely to be important in conserved catalytic or regulatory functions of Nox proteins, although, the detailed biochemical roles of most of these residues are not yet fully understood. The analysis of the *Phox*-related regulatory subunits points to a novel evolutionary history of Nox regulatory proteins. The earliest Nox2 and p47phox- and p22phox-like genes are appearing in the unicellular choanoflagellate before the emergence of the metazoa. Duplication of primordial p47phox- and

A.



B.

AD-like region

Hs-p67phox	KDYLGKATVVASVVDQD	212
Hs-NOXA1	VDFLGKAKVVASAIPDD	213
At-hypo-p67-L1	KVVVMDKEVIASEIVDC	306
At-hypo-p67-L2	KVAVMDKEVIASEIKED	276
At-hypo-p67-L3	TSKEQEKVIEEELLVI	279
At-hypo-p67-L4	SPKGSKVEELGSSSVAV	354

Figure 14**Predicted p67phox-like proteins in higher plants.**

(A) Predicted domain structures of hypothetical p67phox-like proteins (At-hypo-p67-L1 to -L4, Os-hypo-p67-L1) of *A. thaliana* and *O. sativa*, and a fungal *F. graminearum* (Fg-) NOXR are shown, along with their GenBank™ accession numbers. "basic" and "OPCA" in parenthesis indicate six characteristic residues of PB1 domains as shown in Figure 12. "D" and "R" below the TPR region of Fg-NOXR represent two conserved amino acid residues that are important for binding to Rac as indicated in Figure 5. (B) Alignments of the putative AD-like regions are shown (see details in Additional file 17). Boxes indicate lysine and valine residues corresponding to the Lys-196 and Val-204 of human p67phox, respectively. Underlines in alignment indicates amino acid with a hydrophobic side group.

p67phox-like genes occurred in vertebrates, resulting in the emergence of NOXO1-p47phox and NOXA1-p67phox genes, respectively. Supporting this evolutionary origin, the NOXO1 protein in the fish *O. latipes* still retains the p47phox-characteristic "AIR". A predicted motif structure of ancestral p47phox suggests a greatly expanded family of p47phox including SH3PXD2 in the animal lineages. Detailed sequence analyses proposed an evolution model that p40phox originally participated in regulating both Nox1 and Nox2 in early vertebrates, but in mammals the regulation of Nox1 became independent of p40phox. Thus, this report provides keys to understand the evolutionary history of Nox and the regulatory subunits, and provides new information that may be useful in elucidating new molecular details of the activation mechanisms of Nox enzymes including the human orthologs.

Note: While we were in the process of revising this manuscript, a review article about structure and function of the PB1 domain appeared [116]. The review will help to understand the structural features of the PB1 domain that are often mentioned in the present study. In the article, authors also described the presence of the hypothetical genes coding TPR and PB1 domain-containing proteins in the *A. thaliana* genomes and argued about a possibility of *A. thaliana* p67phox.

Methods**Gene identification**

Homologues/orthologs of Nox/Duox and regulatory subunits sequences were assembled from the following species: *H. sapiens* (human), *C. familiaris* (dog), *R. norvegicus* (rat), *M. musculus* (mouse), *G. gallus* (chicken), *X. tropicalis* (frog), *D. rerio* (zebrafish), *T. rubripes* (fugu), *T. nigroviridis* (tetraodon), *O. latipes* (medaka), *C. intestinalis* (ascidian), *S. purpuratus* (sea urchin), *D. pulex* (waterflea), *C. briggsae* (nematode), *C. elegans* (nematode), *N. vectensis* (sea anemone), *L. gigantea* (gastropod snail), *M. brevicollis* (choanoflagellate), *D. discoideum* (slime mold amoeba), *C. merolae* (red alga), *A. nidulans* (fungus-An), *M. grisea* (fungus-Mg), *F. graminearum* (fungus-Fg), *S. cerevisiae* (fungus-Sc), *C. albicans* (fungus-Ca), *Y. lipolytica* (fungus-Yl), *S. pombe* (fungus-Sp), *Coprinopsis cinerea* (*C. cinerea*, fungus-Cc), *L. bicolor* (fungus-Lb), *P. placenta* (fungus-Pp), *Puccinia graminis* (*P. graminis*, fungus-Pg), *P. blakesleeana* (fungus-Pb), *B. dendrobatidis* (fungus-Bd), *P. sojae* (oomycete), *Phaeodactylum tricornutum* (*P. tricornutum*, diatom), *Physcomitrella patens* (*P. patens* moss), *A. thaliana* (land plant), and *O. sativa* (land plant). Using NCBI-HomoloGene [117], existing homologues/orthologs of regulatory subunits were assembled: p47phox, NOXO1, p67phox, NOXA1, p40phox and p22phox orthologs of human, dog, mouse, rat, and p47phox and p67phox and NOXO1 orthologs of chicken. In addition, we assembled amino acid sequences that have been cloned and

reported: p47phox, p67phox, and p22phox of *T. rubripes* [89] and *C. intestinalis* [61]; p67-like protein of *D. discoideum* [56]; NOXR of fungi including *A. nidulans*, *M. grisea*, *F. graminearum* [55,57]. Based on these sequences, BLASTP searches were performed to obtain predicted regulatory subunits, related Nox genes (including genes that has been reported such as *P. sojae* rboh [99], *C. merolae* rboh1/2 [118], *P. tricornutum* rboh1 [118]), new regulator-binding partners, using the NCBI protein database for vertebrates, *C. intestinalis*, *C. briggsae*, *D. discoideum*, *A. thaliana*, *O. sativa* [103], *S. purpuratus* [63], and for fungi (*A. nidulans*, *M. grisea*, *F. graminearum*, *S. cerevisiae*, *C. albicans*, *Y. lipolytica*, *S. pombe*, *C. cinerea*) [119], the Doe Joint Genome Institute database for *L. bicolor*, *P. placenta*, *D. pulex*, *N. vectensis*, *L. gigantea*, *M. brevicollis*, *P. blakesleeanus*, *P. sojae*, *P. tricornutum*, *P. patens* [120], and the Broad Institute FGI database for *P. graminis* and *B. dendrobatidis* [121]. Sequences that had more than 50% identity to the sequence of the closest template were chosen as orthologs or isologs. Assembled sequences, including newly defined, were annotated based on molecular taxonomy analysis of the domains characteristic in each regulator. Characteristic motifs of the regulatory subunits are predicted by the Pfam program [122]. All amino acid sequences and the accession numbers of assembled and analyzed regulator orthologs are listed in Additional material (see Additional files 1, 5, 7, 9, 10, 12, 13, 15, 16, 18).

Phylogenetic analysis and synteny

Multiple sequence alignment and phylogenetic analyses were carried out with ClustalW [123] and trees were reconstructed by the neighbor-joining method as previously described [1]. Amino acid sequences of regulatory subunits were trimmed and aligned to the domains indicated in each figure. Bootstrap values of 1,000 replications are shown at the branches as percentages. Equivalent to 0.1 amino acid substitutions per site indicate evolutionary distances as inferior bars. To elucidate the synteny, we used the Ensembl AlignSliceView program [124]. Conserved marker genes among the genomes of vertebrates are defined (see Additional file 2).

Abbreviations

Nox, NADPH oxidase; Duox, dual oxidase; ROS, reactive oxygen species; TM, transmembrane; NOXO1, Nox Organizer 1; NOXA1, Nox Activator 1; TPR, the tetratricopeptide repeats; AD, activation domain; SH3, Src homology 3; PX, Phox homology domain; PRR, proline-rich region; P-basic, proline-basic region; PB1, Phox/Bem 1; OPCA motif, OPR/PC/AID motif; ADSIS, AD-SH3 Intervening Sequence; SH3PXD2, SH3 and PX domains 2; SRR, serine-rich region; RhoGEF, Rho guanine nucleotide exchange factor; CBS, cystathionine beta-synthase.

Competing interests

The author(s) declares that there are no competing interests.

Authors' contributions

The majority of the work here described was carried out by TK. JDL critically revised the manuscript for important intellectual content and data analysis. Both authors read and approved the final manuscript.

Additional material

Additional file 1

Amino acid sequences of Nox regulatory subunits of 12 deuterostomes, three fungi, and a slime mold amoeba; p47phox, NOXO1, p67phox, NOXA1, NOXR, p40phox, p22phox. Amino acid sequences Nox and Duox proteins of *H. sapiens*, *C. familiaris*, *R. norvegicus*, *M. musculus*, *G. gallus*, *X. tropicalis*, *D. rerio*, *T. rubripes*, *T. nigroviridis*, *O. latipes*, *C. intestinalis*, *S. purpuratus*, *A. nidulans*, *M. grisea*, *F. graminearum* and *D. discoideum* are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S1.doc>]

Additional file 2

Abbreviations of gene names in Figure 2. Names of the marker genes used in Figures 2 are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S2.pdf>]

Additional file 3

Alignment of characterized domains of Nox regulatory subunits.

Alignments of (i) PX and tandem SH3 domains of Nox organizers of deuterostomes, (ii) TPR and activation domains of Nox activators of deuterostomes and fungi (*A. nidulans*, *M. grisea*, *F. graminearum*) and amoeba, (iii) the N-terminal SH3 domain of vertebrate p67phox, (iv) PB1 domain of p67phox and NOXA1 of vertebrates and NOXR of fungi (*A. nidulans*, *M. grisea*, *F. graminearum*) and amoeba, (v) the C-terminal SH3 domain of vertebrate p67phox, vertebrate NOXA1 and *S. purpuratus* p67phox-like protein, (vi) PX domain of vertebrate p40phox, (vii) Proline (P)-basic, SH3 and PB1 domain of vertebrate p40phox.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S3.pdf>]

Additional file 4

Alignment of full-length p22phox (vertebrate p22phox and *C. intestinalis* p22phox-like protein). Alignments of p22phox of chordates.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S4.pdf>]

Additional file 5

Amino acid sequences of Nox/Duox proteins. Amino acid sequences Nox and Duox proteins of the waterflea *D. pulex*, the gastropod snail-limpet *L. gigantea*, the nematode *C. briggsae*, the sea anemone *N. vectensis*, the choanoflagellate *M. brevicollis*, the fungi (*C. cinerea*, *L. bicolor*, *P. placenta*, *P. graminis*, *P. blakesleeana*, and *B. dendrobatidis*), the plant *O. sativa*, and the red alga *C. merolae* are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S5.doc>]

Additional file 6

Molecular taxonomy of Nox and Duox proteins containing newly identified sequences. molecular taxonomy built by amino acid sequences of Nox and Duox proteins of *D. pulex*, *L. gigantea*, *C. briggsae*, *N. vectensis*, *M. brevicollis*, *C. intestinalis*, *S. purpuratus*, and *H. sapiens* is shown.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S6.pdf>]

Additional file 7

Amino acid sequences of putative Nox regulatory subunits of *L. gigantea*, *N. vectensis*, *M. brevicollis*. Amino acid sequences of putative regulatory subunit proteins of *L. gigantea* (*p47phox*-, *p67phox*-, *p22phox*-like proteins), *N. vectensis* (*p47phox*-, *p22phox*-like proteins), and *M. brevicollis* (*p47phox*-, *p22phox*-like proteins) are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S7.doc>]

Additional file 8

Alignments of putative regulatory subunit proteins of *L. gigantea* (*p47phox*-, *p67phox*-, *p22phox*-like proteins), *N. vectensis* (*p47phox*-, *p22phox*-like proteins), and *M. brevicollis* (*p47phox*-, *p22phox*-like proteins). Alignment of *p67phox*-like protein (*L. gigantea* and *H. sapiens*), *p47phox*-like protein (*L. gigantea*, *N. vectensis*, *M. brevicollis*, *S. purpuratus*, and *H. sapiens*), *p22phox*-like proteins (*L. gigantea*, *N. vectensis*, *M. brevicollis*, and *H. sapiens*) are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S8.pdf>]

Additional file 9

Amino acid sequences of Fre of an amoebflagellate. Amino acid sequences of predicted Fre proteins of the amoebflagellate *Naegleria gruberi* are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S9.doc>]

Additional file 10

Amino acid sequences of SH3PXD2 proteins. Amino acid sequences vertebrate and urochordate SH3 and PX domains 2 (SH3PXD2) proteins are provided. Alignments of PX and bis-SH3 domains of SH3PXD2, *p47phox*, *NOXO1*, *p47phox*-like proteins are shown.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S10.doc>]

Additional file 11

Alignment of vertebrate *p47phox*, vertebrate *NOXO1*, *p47phox*-like protein, and chordate *SH3PXD2* proteins. Alignments of *SH3PXD2a* and *2b* of *H. sapiens*, *C. familiaris*, *R. norvegicus*, *M. musculus*, *G. gallus*, *X. tropicalis*, *D. rerio*, *T. rubripes*, *T. nigroviridis*, *O. latipes*, and *C. intestinalis* with other *p47phox* family proteins are shown.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S11.pdf>]

Additional file 12

Amino acid sequences and alignment of predicted fungal NOXR partner: *Bem1*, *Cdc24*, *CBSn-PB1*. Amino acid sequences *Bem1*, *Cdc24*, *CBSn-PB1* of *A. nidulans*, *M. grisea* and *F. graminearum* are provided. Alignments of each gene family are shown.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S12.pdf>]

Additional file 13

Amino acid sequences of NOXR, *Bem1*, *Cdc24*, and *CBSn-PB1* proteins of other fungi than *M. grisea*, *F. graminearum*, and *A. nidulans*. Amino acid sequences *Bem1*, *Cdc24*, and *CBSn-PB1* proteins of *S. cerevisiae*, *C. albicans*, *Y. lipolytica*, *S. pombe*, *C. cinerea*, *L. bicolor*, *P. placenta*, *P. graminis*, *P. blakesleeana*, and *B. dendrobatidis* are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S13.doc>]

Additional file 14

Alignments of fungal NOXR, *Bem1*, *Cdc24*, and *CBSn-PB1* amino acid sequences. Alignments of fungal NOXR, *Bem1*, *Cdc24*, and *CBSn-PB1* of *A. nidulans*, *M. grisea*, *F. graminearum*, *S. cerevisiae*, *C. albicans*, *Y. lipolytica*, *S. pombe*, *C. cinerea*, *L. bicolor*, *P. placenta*, *P. graminis*, *P. blakesleeana*, and *B. dendrobatidis* are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S14.pdf>]

Additional file 15

Amino acid sequences of PB1 domain-containing proteins of *D. discoideum* and alignment of PB1 domains. Amino acid sequences of PB1 domain-containing proteins of the slime mold amoeba *D. discoideum* (*Dd-PB1-A* to *-E*) and an alignment of the PB1 domains are provided.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S15.pdf>]

Additional file 16

Amino acid sequences of hypothetical *p67phox*-like (*hypo-p67-L*) of *Arabidopsis thaliana* and *Oryza sativa*. Sequences of *A. thaliana* and *O. sativa* proteins that contain the TPR and PB1 domains (*At-hypo-p67-L1* to *L4* and *Or-hypo-p67-L1*) are shown.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S16.doc>]

Additional file 17

Alignment of hypothetical p67phox-like (hypo-p67-L) of *Arabidopsis thaliana* and *Oryza sativa*. (i) Alignment of four hypo-p67-L1 to -L4 of *A. thaliana*, (ii) comparison of TPR and AD domain between the *A. thaliana* and *O. sativa* proteins (At-hypo-p67-L1-4 and Os-hypo-p67-L1) and human p67phox (1–212 residues) and NOXA1 (1–213 residues) proteins are shown.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S17.pdf>]

Additional file 18

ID numbers of hypothetical PB1 domain-containing genes of *Arabidopsis thaliana*, except for hypo-p67-L genes. a total of 27 ID numbers of *Arabidopsis thaliana* genes are shown.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2148-7-178-S18.pdf>]

Acknowledgements

We express our appreciation to Mrs. Heather Jackson for reading and commenting on the manuscript and to Dr. Yasushi Okamura for suggesting sea urchin genome research. We thank Drs. Seiji Takeda and Takehiko Ueyama for commenting about *A. thaliana* p67phox-like genes and a conserved lysine residue in AD of p67phox, respectively. We also thank anonymous reviewers for suggestive comments to improve content of the manuscript. We are grateful to Mrs. Asami Kawahara for great cooperation and patience during long nights and weekends of endless computational analysis at home. This work was supported by NIH grants CA105116 and GM067717.

References

- Kawahara T, Quinn MT, Lambeth JD: **Molecular evolution of the reactive oxygen-generating NADPH oxidase (Nox/Duox) family of enzymes.** *BMC Evolutionary Biology* 2007, **7**(109):.
- Bedard K, Lardy B, Krause KH: **NOX family NADPH oxidases: Not just in mammals.** *Biochimie* 2007, **89**(9):1107-1112.
- Vignais PV: **The superoxide-generating NADPH oxidase; structural aspects and activation mechanism.** *Cellular and Molecular Life Sciences* 2002, **59**(9):1428-1459.
- Heyworth PG, Cross AR, Curnutte JT: **Chronic granulomatous disease.** *Current Opinion in Immunology* 2003, **15**(5):578-584.
- Lambeth JD: **Nox enzymes, ROS, and chronic disease: an example of antagonistic pleiotropy.** *Free Radical Biology and Medicine* 2007, **43**(3):332-347.
- Nauseef WM: **Assembly of the phagocyte NADPH oxidase.** *Histochemistry and Cell Biology* 2004, **122**(4):277-291.
- Geiszt M: **NADPH oxidases: new kids on the block.** *Cardiovascular Research* 2006, **71**(2):289-299.
- Leto TL, Geiszt M: **Role of Nox family NADPH oxidases in host defense.** *Antioxidants & Redox Signaling* 2006, **8**(9-10):1549-1561.
- Lambeth JD: **NOX enzymes and the biology of reactive oxygen.** *Nature Review Immunology* 2004, **4**(3):181-189.
- Sumimoto H, Miyano K, Takeya R: **Molecular composition and regulation of the Nox family NAD(P)H oxidases.** *Biochemical and Biophysical Research Communications* 2005, **38**(1):677-686.
- Lambeth JD, Kawahara T, Diebold B: **Regulation of Nox and Duox enzymatic activity and expression.** *Free Radical Biology and Medicine* 2007, **43**(3):319-331.
- Ishibashi F, Nunoi H, Endo F, Matsuda I, Kanegasaki S: **Statistical and mutational analysis of chronic granulomatous disease in Japan with special reference to gp91-phox and p22-phox deficiency.** *Human Genetics* 2000, **106**(5):473-481.
- Bionda C, Li XJ, van Bruggen R, Eppink M, Roos D, Morel F, Stasia MJ: **Functional analysis of two-amino acid substitutions in gp91 phox in a patient with X-linked flavocytochrome b558-positive chronic granulomatous disease by means of transgenic PLB-985 cells.** *Human Genetics* 2004, **115**(5):418-427.
- Forbes L, Truong O, Wientjes FB, Moss SJ, Segal AW: **The major phosphorylation site of the NADPH oxidase component p67phox is Thr233.** *Biochemical Journal* 1999, **338**(1):99-105.
- Regier DS, Greene DG, Sergeant S, Jesaitis AJ, McPhail LC: **Phosphorylation of p22phox is mediated by phospholipase D-dependent and -independent mechanisms. Correlation of NADPH oxidase activity and p22phox phosphorylation.** *Journal of Biological Chemistry* 2000, **275**(37):28406-28412.
- Bouin AP, Grandvaux N, Vignais PV, Fuchs A: **p40(phox) is phosphorylated on threonine 154 and serine 315 during activation of the phagocyte NADPH oxidase. Implication of a protein kinase c-type kinase in the phosphorylation process.** *Journal of Biological Chemistry* 1998, **273**(46):30097-30103.
- Knaus UG, Heyworth PG, Evans T, Curnutte JT, Bokoch GM: **Regulation of phagocyte oxygen radical production by the GTP-binding protein Rac 2.** *Science* 1991, **254**(5037):1512-1515.
- Kami K, Takeya R, Sumimoto H, Kohda D: **Diverse recognition of non-PxxP peptide ligands by the SH3 domains from p67(phox), Grb2 and Pex13p.** *The EMBO Journal* 2002, **21**(16):4268-4276.
- El Benna J, Faust L, Babior B: **The phosphorylation of the respiratory burst oxidase component p47phox during neutrophil activation.** *Journal of Biological Chemistry* 1994, **269**:23431-23436.
- Ago T, Nunoi H, Ito T, Sumimoto H: **Mechanism for phosphorylation-induced activation of the phagocyte NADPH oxidase protein p47(phox). Triple replacement of serines 303, 304, and 328 with aspartates disrupts the SH3 domain-mediated intramolecular interaction in p47(phox), thereby activating the oxidase.** *Journal of Biological Chemistry* 1999, **274**(47):33644-33653.
- Groemping Y, Lapouge K, Smerdon SJ, Rittinger K: **Molecular basis of phosphorylation-induced activation of the NADPH oxidase.** *Cell* 2003, **113**(3):343-355.
- Rotrosen D, Leto TL: **Phosphorylation of neutrophil 47-kDa cytosolic oxidase factor. Translocation to membrane is associated with distinct phosphorylation events.** *Journal of Biological Chemistry* 1990, **265**(32):19910-19915.
- Ago T, Takeya R, Hiroaki H, Kuribayashi F, Ito T, Kohda D, Sumimoto H: **The PX domain as a novel phosphoinositide-binding module.** *Biochemical & Biophysical Research Communications* 2001, **287**(3):733-738.
- Kanai F, Liu H, Field SJ, Akbary H, Matsuo T, Brown GE, Cantley LC, Yaffe MB: **The PX domains of p47phox and p40phox bind to lipid products of PI(3)K.** *Nature Cell Biology* 2001, **3**(7):675-678.
- Karathanassis D, Stahelin RV, Bravo J, Perisic O, Pacold CM, Cho W, Williams RL: **Binding of the PX domain of p47(phox) to phosphatidylinositol 3,4-bisphosphate and phosphatidic acid is masked by an intramolecular interaction.** *The EMBO Journal* 2002, **21**(19):5057-5068.
- Ligeti E, Tardif M, Vignais PV: **Activation of O2- generating oxidase of bovine neutrophils in a cell-free system. Interaction of a cytosolic factor with the plasma membrane and control by G nucleotides.** *Biochemistry* 1989, **28**:7116-7123.
- Koga H, Terasawa H, Nunoi H, Takeshige K, Inagaki F, Sumimoto H: **Tetratricopeptide repeat (TPR) motifs of p67phox participate in interaction with the small GTPase Rac and activation of the phagocyte NADPH oxidase.** *Journal of Biological Chemistry* 1999, **274**:25051-25060.
- Lapouge K, Smith SJ, Walker PA, Gamblin SJ, Smerdon SJ, Rittinger K: **Structure of the TPR domain of p67phox in complex with Rac.GTP.** *Molecular Cell* 2000, **6**(4):899-907.
- Sarfstein R, Gorzalczyk Y, Mizrahi A, Berdichevsky Y, Molshanski-Mor S, Weinbaum C, Hirshberg M, Dagher MC, Pick E: **Dual role of Rac in the assembly of NADPH oxidase, tethering to the membrane and activation of p67phox: a study based on mutagenesis of p67phox-Rac1 chimeras.** *Journal of Biological Chemistry* 2004, **279**(16):16007-16016.
- Han C-H, Freeman JLR, Lee T, Motalebi SA, Lambeth JD: **Regulation of the neutrophil respiratory burst oxidase: Identification of an activation domain in p67phox.** *Journal of Biological Chemistry* 1998, **273**:16663-16668.
- Nisimoto Y, Motalebi S, Han C-H, Lambeth JD: **The p67phox activation domain regulates electron transfer flow from**

- NADPH to flavin in flavocytochrome b_{558} .** *Journal of Biological Chemistry* 1999, **274**:22999-23005.
32. Wilson MI, Gill DJ, Perisic O, Quinn MT, Williams RL: **PBI domain-mediated heterodimerization in NADPH oxidase and signaling complexes of atypical protein kinase C with Par6 and p62.** *Molecular Cell* 2003, **12**(1):39-50.
 33. Kuribayashi F, Nunoi H, Wakamatsu K, Tsunawaki S, Sato K, Ito T, Sumimoto H: **The adaptor protein p40(phox) as a positive regulator of the superoxide-producing phagocyte oxidase.** *The EMBO Journal* 2002, **21**(23):6312-6320.
 34. Nakamura R, Sumimoto H, Mizuki K, Hata K, Ago T, Kitajima S, Takeshige K, Sakaki Y, Ito T: **The PC motif: a novel and evolutionarily conserved sequence involved in interaction between p40phox and p67phox, SH3 domain-containing cytosolic factors of the phagocyte NADPH oxidase.** *European Journal of Biochemistry* 1998, **251**(3):583-589.
 35. Ellison CD, Davidson K, Ferguson GJ, O'Connor R, Stephens LR, Hawkins PT: **Neutrophils from p40phox-/- mice exhibit severe defects in NADPH oxidase regulation and oxidant-dependent bacterial killing.** *Journal of Experimental Medicine* 2006, **203**(8):1927-1937.
 36. Suh YA, Arnold RS, Lassegue B, Shi J, Xu X, Sorescu D, Chung AB, Griendling KK, Lambeth JD: **Cell transformation by the superoxide-generating oxidase Mox1.** *Nature* 1999, **401**(6748):79-82.
 37. Bedard K, Krause KH: **The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology.** *Physiological Reviews* 2007, **87**(1):245-313.
 38. Banfi B, Clark RA, Steger K, Krause KH: **Two novel proteins activate superoxide generation by the NADPH oxidase NOX1.** *Journal of Biological Chemistry* 2003, **278**(6):3510-3513.
 39. Geiszt M, Lekstrom K, Witta J, Leto TL: **Proteins Homologous to p47phox and p67phox Support Superoxide Production by NAD(P)H Oxidase 1 in Colon Epithelial Cells.** *Journal of Biological Chemistry* 2003, **278**(22):20006-20012.
 40. Takeya R, Ueno N, Kami K, Taura M, Kohjima M, Izaki T, Nunoi H, Sumimoto H: **Novel human homologues of p47phox and p67phox participate in activation of superoxide-producing NADPH oxidases.** *Journal of Biological Chemistry* 2003, **278**(27):25234-25246.
 41. Cheng G, Lambeth JD: **NOXO1, Regulation of Lipid Binding, Localization, and Activation of Nox1 by the Phox Homology (PX) Domain.** *Journal of Biological Chemistry* 2004, **279**(6):4737-4742.
 42. Ueyama T, Geiszt M, Leto TL: **Involvement of Rac1 in activation of multicompartment Nox1- and Nox3-based NADPH oxidases.** *Molecular and Cellular Biology* 2006, **26**(6):2160-2174.
 43. Kawahara T, Kohjima M, Kuwano Y, Mino H, Teshima-Kondo S, Takeya R, Tsunawaki S, Wada A, Sumimoto H, Rokutan K: **Helicobacter pylori lipopolysaccharide activates Rac1 and transcription of NADPH oxidase Nox1 and its organizer NOXO1 in guinea pig gastric mucosal cells.** *American Journal of Physiology - Cell Physiology* 2005, **288**(2):C450-457.
 44. Cheng G, Diebold BA, Hughes Y, Lambeth JD: **Nox1-dependent reactive oxygen generation is regulated by Rac1.** *Journal of Biological Chemistry* 2006, **281**(26):17718-17726.
 45. Miyano K, Ueno N, Takeya R, Sumimoto H: **Direct involvement of the small GTPase Rac in activation of the superoxide-producing NADPH oxidase Nox1.** *Journal of Biological Chemistry* 2006, **281**(31):21857-21868.
 46. Kawahara T, Kuwano Y, Teshima-Kondo S, Takeya R, Sumimoto H, Kishi K, Tsunawaki S, Hirayama T, Rokutan K: **Role of nicotinamide adenine dinucleotide phosphate oxidase 1 in oxidative burst response to Toll-like receptor 5 signaling in large intestinal epithelial cells.** *The Journal of Immunology* 2004, **172**(5):3051-3058.
 47. Yamamoto A, Kami K, Takeya R, Sumimoto H: **Interaction between the SH3 domains and C-terminal proline-rich region in NADPH oxidase organizer 1 (Noxo1).** *Biochemical and Biophysical Research Communications* 2007, **352**:560-565.
 48. Banfi B, Malgrange B, Knisz J, Steger K, Dubois-Dauphin M, Krause KH: **NOX3, a superoxide-generating NADPH oxidase of the inner ear.** *Journal of Biological Chemistry* 2004, **279**(44):46065-46072.
 49. Cheng G, Ritsick DR, Lambeth JD: **Nox3 regulation by NOXO1, p47phox and p67phox.** *Journal of Biological Chemistry* 2004.
 50. Kiss PJ, Knisz J, Zhang Y, Baltrusaitis J, Sigmund CD, Thalmann R, Smith RJ, Verpy E, Banfi B: **Inactivation of NADPH oxidase organizer 1 results in severe imbalance.** *Current Biology* 2006, **16**(2):208-213.
 51. Ueno N, Takeya R, Miyano K, Kikuchi H, Sumimoto H: **The NADPH oxidase Nox3 constitutively produces superoxide in a p22phox-dependent manner: its regulation by oxidase organizers and activators.** *Journal of Biological Chemistry* 2005, **280**(24):23328-23339.
 52. Ambasta RK, Kumar P, Griendling KK, Schmidt HH, Busse R, Brandes RP: **Direct interaction of the novel Nox proteins with p22phox is required for the formation of a functionally active NADPH oxidase.** *Journal of Biological Chemistry* 2004, **279**(44):45935-45941.
 53. Kawahara T, Ritsick D, Cheng G, Lambeth JD: **Point mutations in the proline-rich region of p22phox are dominant inhibitors of Nox1- and Nox2-dependent reactive oxygen generation.** *Journal of Biological Chemistry* 2005, **280**(36):31859-31869.
 54. Martyn K, Frederick LM, von Loehneysen K, Dinuer MC, Knaus UG: **Functional analysis of Nox4 reveals unique characteristics compared to other NADPH oxidases.** *Cellular Signalling* 2006, **18**(1):69-82.
 55. Takemoto D, Tanaka A, Scott B: **NADPH oxidases in fungi: Diverse roles of reactive oxygen species in fungal cellular differentiation.** *Fungal Genetics and Biology* 2007.
 56. Lardy B, Bof M, Aubry L, Paclet M, Morel F, Satre M, Klein G: **NADPH oxidase homologs are required for normal cell differentiation and morphogenesis in Dictyostelium discoideum.** *Biochimica et Biophysica Acta* 2005, **1744**(2):199-212.
 57. Takemoto D, Tanaka A, Scott B: **A p67Phox-like regulator is recruited to control hyphal branching in a fungal-grass mutualistic symbiosis.** *Plant Cell* 2006, **18**(10):2807-2821.
 58. Aparicio S, Chapman J, Stupka E, Putnam N, Chia JM, Dehal P, Christoffels A, Rash S, Hoon S, Smit A, et al.: **Whole-genome shotgun assembly and analysis of the genome of Fugu rubripes.** *Science* 2002, **297**(5585):1301-1310.
 59. **Animal-Genome-Size-Database** [<http://http://www.genome-size.com/>]
 60. Jaillon O, Aury JM, Brunet F, Petit JL, Stange-Thomann N, Mauceli E, Bouneau L, Fischer C, Ozouf-Costaz C, Bernot A: **Genome duplication in the teleost fish Tetraodon nigroviridis reveals the early vertebrate proto-karyotype.** *Nature* 2004, **431**(7011):946-957.
 61. Inoue Y, Ogasawara M, Moroi T, Satake M, Azumi K, Morimoto T, Nakanishi T: **Characteristics of NADPH oxidase genes (Nox2, p22, p47, and p67) and Nox4 gene expressed in blood cells of juvenile Ciona intestinalis.** *Immunogenetics* 2005, **57**(7):520-534.
 62. Sea Urchin Genome Sequencing Consortium, Sodergren EWG, Davidson EH, Cameron RA, Gibbs RA, Angerer RC, Angerer LM, Arnott MI, Burgess DR, Burke RD, Coffman JA, Dean M, Elphick MR, Ettensohn CA, Foltz KR, Hamdoun A, Hynes RO, Klein WH, Marzluff W, McClay DR, Morris RL, Mushegian A, Rast JP, Smith LC, Thorndyke MC, Vacquier VD, Wessel GM, Wray G, Zhang L, Elisk CG, Ermo-laeva O, Hlavina W, Hofmann G, Kitts P, Landrum MJ, Mackey AJ, Maglott D, Panopoulou G, Poustka AJ, Pruitt K, Sapozhnikov V, Song X, Souvorov A, Solovyev V, Wei Z, Whittaker CA, Worley K, Durbin KJ, Shen Y, Fedrigo O, Garfield D, Haygood R, Primus A, Satija R, Severson T, Gonzalez-Garay ML, Jackson AR, Milosavljevic A, Tong M, Killian CE, Livingston BT, Wilt FH, Adams N, Belle R, Carbonneau S, Cheung R, Cormier P, Cosson B, Croce J, Fernandez-Guerra A, Genevieve AM, Goel M, Kelkar H, Morales J, Mulner-Lorillon O, Robertson AJ, Goldstone JV, Cole B, Epel D, Gold B, Hahn ME, Howard-Ashby M, Scally M, Stegeman JJ, Allgood EL, Cool J, Judkins KM, McCafferty SS, Musante AM, Obar RA, Rawson AP, Rossetti BJ, Gibbons IR, Hoffman MP, Leone A, Istrail S, Materna SC, Samanta MP, Stolc V, Tongprasit W, Tu Q, Bergeron KF, Brandhorst BP, Whittle J, Berney K, Bottjer DJ, Calestani C, Peterson K, Chow E, Yuan QA, Elhaik E, Graur D, Reese JT, Bosdet I, Heesun S, Marra MA, Schein J, Anderson MK, Brockton V, Buckley KM, Cohen AH, Fugmann SD, Hibino T, Loza-Coll M, Majeske AJ, Messier C, Nair SV, Pancer Z, Terwilliger DP, Agca C, Arboleda E, Chen N, Churcher AM, Hallbook F, Humphrey GW, Idris MM, Kiyama T, Liang S, Mellott D, Mu X, Murray G, Olinski RP, Raible F, Rowe M, Taylor JS, Tessmar-Raible K, Wang D, Wilson KH, Yaguchi S, Gaasterland T, Galindo BE, Gunaratne HJ, Juliano C, Kinukawa M, Moay GW, Neill AT, Nomura M, Rausch M, Reade A, Roux MM, Song JL, Su YH, Townley IK, Voron-

- ina E, Wong JL, Amore G, Branno M, Brown ER, Cavaliere V, Duboc V, Duloquin L, Flytzanis C, Gache C, Lapraz F, Lepage T, Locascio A, Martinez P, Matassi G, Matranga V, Range R, Rizzo F, Rottinger E, Beane W, Bradham C, Byrum C, Glenn T, Hussain S, Manning G, Miranda E, Thomason R, Walton K, Wikramanayake A, Wu SY, Xu R, Brown CT, Chen L, Gray RF, Lee PY, Nam J, Oliveri P, Smith J, Muzny D, Bell S, Chacko J, Cree A, Curry S, Davis C, Dinh H, Dugan-Rocha S, Fowler J, Gill R, Hamilton C, Hernandez J, Hines S, Hume J, Jackson L, Jolivet A, Kovar C, Lee S, Lewis L, Miner G, Morgan M, Nazareth LV, Okwuonu G, Parker D, Pu LL, Thorn R, Wright R: **The genome of the sea urchin *Strongylocentrotus purpuratus***. *Science* 2006, **314**(5801):941-952.
63. **NCBI-Sea-Urchin-Genome-Database** [<http://www.ncbi.nlm.nih.gov/genome/seq/BlastGen/BlastGen.cgi?taxid=7668>]
 64. Hoegg S, Meyer A: **Hox clusters as models for vertebrate genome evolution**. *Trends in Genetics* 2005, **21**(8):421-424.
 65. Takeya R, Taura M, Yamasaki T, Naito S, Sumimoto H: **Expression and function of Noxo I gamma, an alternative splicing form of the NADPH oxidase organizer I**. *FEBS Journal* 2006, **273**(16):3663-3677.
 66. Hata K, Ito T, Takeshige K, Sumimoto H: **Anionic amphiphile-independent activation of the phagocyte NADPH oxidase in a cell-free system by p47^{phox} and p67^{phox} both in C terminally truncated forms**. *Journal of Biological Chemistry* 1998, **273**:4232-4236.
 67. Nagasawa T, Ebisu K, Inoue Y, Miyano K, Tamura M: **A new role of Pro-73 of p47^{phox} in the activation of neutrophil NADPH oxidase**. *Archives of Biochemistry and Biophysics* 2003, **416**(1):92-100.
 68. Ogura K, Nobuhisa I, Yuzawa S, Takeya R, Torikai S, Saikawa K, Sumimoto H, Inagaki F: **NMR solution structure of the tandem Src homology 3 domains of p47^{phox} complexed with a p22^{phox}-derived proline-rich peptide**. *Journal of Biological Chemistry* 2006, **281**(6):3660-3668.
 69. Ago T, Kuribayashi F, Hiroaki H, Takeya R, Ito T, Kohda D, Sumimoto H: **Phosphorylation of p47^{phox} directs phox homology domain from SH3 domain toward phosphoinositides, leading to phagocyte NADPH oxidase activation**. *Proceedings of the National Academy of Sciences of the United States of America* 2003, **100**(8):4474-4479.
 70. Finan P, Shimizu Y, Gout I, Hsuan J, Truong O, Butcher C, Bennett P, Waterfield MD, Kellie S: **An SH3 domain and proline-rich sequence mediate an interaction between two components of the phagocyte NADPH oxidase complex**. *Journal of Biological Chemistry* 1994, **269**(19):13752-13755.
 71. Leusen JH, Fluiter K, Hilarius PM, Roos D, Verhoeven AJ, Bolscher BG: **Interactions between the cytosolic components p47^{phox} and p67^{phox} of the human neutrophil NADPH oxidase that are not required for activation in the cell-free system**. *Journal of Biological Chemistry* 1995, **270**(19):11216-11221.
 72. Mizuki K, Takeya R, Kuribayashi F, Nobuhisa I, Kohda D, Nunoi H, Takeshige K, Sumimoto H: **A region C-terminal to the proline-rich core of p47^{phox} regulates activation of the phagocyte NADPH oxidase by interacting with the C-terminal SH3 domain of p67^{phox}**. *Archives of Biochemistry and Biophysics* 2005, **444**(2):185-194.
 73. Curwen V, Eyraas E, Andrews TD, Clarke L, Mongin E, Searle SM, Clamp M: **The Ensembl automatic gene annotation system**. *Genome Research* 2004, **14**(5):942-950.
 74. Diebold BA, Bokoch GM: **Molecular basis for Rac2 regulation of phagocyte NADPH oxidase**. *Nature Immunology* 2001, **2**(3):211-215.
 75. Leto T, Adams AG, de Mendez I: **Assembly of the phagocyte NADPH oxidase: binding of Src homology 3 domains to proline-rich targets**. *The Proceedings of the National Academy of Sciences Online of the United States of America* 1994, **91**(22):10650-10654.
 76. Ito T, Matsui Y, Ago T, Ota K, Sumimoto H: **Novel modular domain PBI recognizes PC motif to mediate functional protein-protein interactions**. *The EMBO Journal* 2001, **20**(15):3938-3946.
 77. Gauss KA, Bunger PL, Siemsen DW, Young CJ, Nelson-Overton L, Prigge JR, Swain SD, Quinn MT: **Molecular analysis of the bison phagocyte NADPH oxidase: cloning and sequencing of five NADPH oxidase cDNAs**. *Comparative Biochemistry & Physiology Part B, Biochemistry & Molecular Biology* 2002, **133**(1):1-12.
 78. Gauss KA, Mascolo PL, Siemsen DW, Nelson LK, Bunger PL, Pagano PJ, Quinn MT: **Cloning and sequencing of rabbit leukocyte NADPH oxidase genes reveals a unique p67(phox) homolog**. *Journal of Leukocyte Biology* 2002, **71**(2):319-328.
 79. Zhan Y, Virbasius JV, Song X, Pomerleau DP, Zhou GW: **The p40^{phox} and p47^{phox} PX domains of NADPH oxidase target cell membranes via direct and indirect recruitment by phosphoinositides**. *Journal of Biological Chemistry* 2002, **277**(6):4512-4518.
 80. Stahelin RV, Burian A, Bruzik KS, Murray D, Cho W: **Membrane binding mechanisms of the PX domains of NADPH oxidase p40^{phox} and p47^{phox}**. *Journal of Biological Chemistry* 2003, **278**(16):14469-14479.
 81. Ueyama T, Tatsuno T, Kawasaki T, Tsujibe S, Shirai Y, Sumimoto H, Leto T, Saito N: **A Regulated Adaptor Function of p40^{phox}: Distinct p67^{phox} Membrane Targeting by p40^{phox} and by p47^{phox}**. *Molecular and Cellular Biology* 2007, **18**(2):441-454.
 82. Honbou K, Minakami R, Yuzawa S, Takeya R, Suzuki NN, Kamakura S, Sumimoto H, Inagaki F: **Full-length p40^{phox} structure suggests a basis for regulation mechanism of its membrane binding**. *EMBO Journal* 2007, **26**(4):1176-1186.
 83. Taylor R, Burritt J, Baniulis D, Foubert T, Lord C, Dinanuer MC, Parkos CA, Jesaitis AJ: **Site-specific inhibitors of NADPH oxidase activity and structural probes of flavocytochrome b: characterization of six monoclonal antibodies to the p22^{phox} subunit**. *The Journal of Immunology* 2004, **173**(12):7349-7357.
 84. Varnai P, Thyagarajan B, Rohacs T, Balla T: **Rapidly inducible changes in phosphatidylinositol 4,5-bisphosphate levels influence multiple regulatory functions of the lipid in intact living cells**. *Journal of Cell Biology* 2006, **175**(3):377-382.
 85. Heo WD, Inoue T, Park WS, Kim ML, Park BO, Wandless TJ, Meyer T: **PI(3,4,5)P3 and PI(4,5)P2 lipids target proteins with polybasic clusters to the plasma membrane**. *Science* 2006, **314**(5804):1458-1461.
 86. Ueyama T, Eto M, Kami K, Tatsuno T, Kobayashi T, Shirai Y, Lennartz MR, Takeya R, Sumimoto H, Saito N: **Isoform-specific membrane targeting mechanism of Rac during Fc gamma R-mediated phagocytosis: positive charge-dependent and independent targeting mechanism of Rac to the phagosome**. *Journal of Immunology* 2005, **175**(4):2381-2390.
 87. **PROSITE-database** [<http://ca.expasy.org/prosite/>]
 88. Suh CI, Stull ND, Li XJ, Tian WH, Price MO, Grinstein S, Yaffe MB, Atkinson S, Dinanuer MC: **The phosphoinositide-binding protein p40^{phox} activates the NADPH oxidase during Fc gamma IIA receptor-induced phagocytosis**. *Journal of Experimental Medicine* 2006, **203**(8):1915-1925.
 89. Inoue Y, Suenaga Y, Yoshiura Y, Moritomo T, Ototake M, Nakanishi T: **Molecular cloning and sequencing of Japanese pufferfish (Takifugu rubripes) NADPH oxidase cDNAs**. *Developmental & Comparative Immunology* 2004, **28**(9):911-925.
 90. Halanych KM: **The new view of animal phylogeny**. *Annual Review of Ecology, Evolution, and Systematics* 2004, **35**:229-256.
 91. Li S: **Specificity and versatility of SH3 and other proline-recognition domains: structural basis and implication for cellular signal transduction**. *Biochemical Journal* 2005, **390**:641-653.
 92. Miller DJ, Hemmrich G, Ball EE, Hayward DC, Khatunin K, Funayama N, Agata K, Bosch TCG: **The innate immune repertoire in Cnidaria-ancestral complexity and stochastic gene loss**. *Genome Biology* 2007, **8**(4):R59.51-R59.13.
 93. James-Clark H: **Note on the infusoria flagellata and the spongiae ciliatae**. *American Journal of Science* 1866, **1**:113-114.
 94. Leadbeater BS, Kelly M: **Evolution of animals-choanoflagellates and sponges**. *Water and Atmosphere* 2001, **9**:9-11.
 95. Snell EA, Furlong RF, Holland PWH: **Hsp70 sequences indicate that choanoflagellates are closely related to animals**. *Current Biology* 2001, **11**:967-970.
 96. King N, Carroll SB: **A receptor tyrosine kinase from choanoflagellates: molecular insights into early animal evolution**. *Proceedings of the National Academy of Sciences of the United States of America* 2001, **98**(26):15032-15037.
 97. King N, Hittinger CT, Carroll SB: **Evolution of key cell signaling and adhesion protein family predates animal origins**. *Science* 2003, **301**:361-363.
 98. King N: **The unicellular ancestry of animal development**. *Developmental Cell* 2004, **7**:313-325.
 99. Lalucque H, Silar P: **NADPH oxidase: an enzyme for multicellularity**. *TRENDS in Microbiology* 2003, **11**(1):9-12.

100. Cavalier-Smith T: **Only six Kingdoms of life.** *Proceedings Royal Society of London B* 2004, **271**:1251-1262.
101. Seals DF, Azucena EF Jr, Pass I, Tesfay L, Gordon R, Woodrow M, Resau JH, Courtneidge SA: **The adaptor protein Tks5/Fish is required for podosome formation and function, and for the protease-driven invasion of cancer cells.** *Cancer Cell* 2005, **7**(2):155-165.
102. Malinin NL, Wright S, Seubert P, Schenk D, Griswold-Prenner I: **Amyloid-beta neurotoxicity is mediated by FISH adapter protein and ADAM12 metalloprotease activity.** *Proceedings of the National Academy of Sciences of the United States of America* 2005, **102**(8):3058-3063.
103. NCBI-BLAST [<http://http://www.ncbi.nlm.nih.gov/BLAST/>]
104. Stahelin RV, Karathanassis D, Murray D, Williams RL, Cho W: **Structural and Membrane Binding Analysis of the Phox Homology Domain of Bem1p: BASIS OF PHOSPHATIDYLINOSITOL 4-PHOSPHATE SPECIFICITY.** *Journal of Biological Chemistry* 2007, **282**(35):25737-25747.
105. France YE, Boyd C, Coleman J, Novick PJ: **The polarity-establishment component Bem1p interacts with the exocyst complex through the Sec15p subunit.** *Journal of Cell Science* 2006, **119**(Pt 5):876-888.
106. Ignoul S, Eggermont J: **CBS domains: structure, function, and pathology in human proteins.** *American Journal of Physiology - Cell Physiology* 2005, **289**(6):C1369-1378.
107. Scott JW, Hawley SA, Green KA, Anis M, Stewart G, Scullion GA, Norman DG, Hardie DG: **CBS domains form energy-sensing modules whose binding of adenosine ligands is disrupted by disease mutations.** *The Journal of Clinical Investigation* 2004.
108. Chung CY, Reddy TB, Zhou K, Firtel RA: **A novel, putative MEK kinase controls developmental timing and spatial patterning in Dictyostelium and is regulated by ubiquitin-mediated protein degradation.** *Genes and Development* 1998, **12**(22):3564-3578.
109. Kobayashi M, Ohura I, Kawakita K, Yokota N, Fujiwara M, Shimamoto K, Doke N, Yoshioka H: **Calcium-dependent protein kinases regulate the production of reactive oxygen species by potato NADPH oxidase.** *Plant Cell* 2007, **19**(3):1065-1080.
110. Jagnandan D, Church JE, Banfi B, Stuehr DJ, Marrero MB, Fulton DJ: **Novel mechanism of activation of NADPH oxidase 5. calcium sensitization via phosphorylation.** *Journal of Biological Chemistry* 2007, **282**(9):6469-6507.
111. Serrander L, Jaquet V, Bedard K, Plastre O, Hartley O, Arnaudeau S, Demaurex N, Schlegel W, Krause KH: **NOX5 is expressed at the plasma membrane and generates superoxide in response to protein kinase C activation.** *Biochimie* 2007, **89**(9):1159-1167.
112. Carol RJ, Dolan L: **The role of reactive oxygen species in cell growth: lessons from root hairs.** *Journal of Experimental Botany* 2006, **57**(8):1829-1834.
113. Keller T, Damude HG, Werner D, Doerner P, Dixon RA, Lamb C: **A plant homolog of the neutrophil NADPH oxidase gp91phox subunit gene encodes a plasma membrane protein with Ca2+ binding motifs.** *Plant Cell* 1998, **10**(2):255-266.
114. Groom QJ, Torres MA, Fordham-Skelton AP, Hammond-Kosack KE, Robinson NJ, Jones JD: **rbohA, a rice homologue of the mammalian gp91phox respiratory burst oxidase gene.** *The Plant Journal* 1996, **3**:515-22.
115. TIGR-Arabidopsis-thaliana-Protein-Database [<http://tigrblast.tigr.org/er-blast/index.cgi?project=ath1>]
116. Sumimoto H, Kamakura S, Ito T: **Structure and function of the PBI domain, a protein interaction module conserved in animals, fungi, amoebas, and plants.** *Sci STKE* 2007, **401**.
117. NCBI-Homologene [<http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=homologene>]
118. Herve C, Tonon T, Collen J, Corre E, Boyen C: **NADPH oxidases in Eukaryotes: red algae provide new hints!** *Current Genetics* 2006, **49**(3):190-204.
119. NCBI-Fungi-Genomes-BLAST-server [http://www.ncbi.nlm.nih.gov/sutils/genom_table.cgi?organism=fungi]
120. JGI-Genomes-Database [http://genome.jgi-psf.org/euk_home.html]
121. Broad-Institute-Genomes-database [http://www.broad.mit.edu/cgi-bin/annotation/fgi/blast_page.cgi]
122. Pfam-search [<http://www.sanger.ac.uk/Software/Pfam/>]
123. ClustalW [<http://http://www.ddbj.nig.ac.jp/search/clustalw-i.html>]
124. Ensembl-AlignSliceView-program [http://www.ensembl.org/Homo_sapiens/alignsliceview]
125. Hedges SB: **The origin and evolution of model organisms.** *Nature Reviews Genetics* 2002, **3**(11):838-849.
126. Sumimoto H, Hata K, Mizuki K, Ito T, Kage Y, Sakaki Y, Fukumaki Y, Nakamura M, Takeshige K: **Assembly and activation of the phagocyte NADPH oxidase: Specific interaction of the N-terminal Src homology 3 domain of p47phox with p22phox is required for activation of the NADPH oxidase complex.** *Journal of Biological Chemistry* 1996, **36**:22152-22158.
127. Sumimoto H, Kage Y, Nunoi H, Sasaki H, Nose T, Fukumaki Y, Ohno M, Minakami S, Takeshige K: **Role of Src homology 3 domains in assembly and activation of the phagocyte NADPH oxidase.** *Proceedings of the National Academy of Sciences of the United States of America* 1994, **91**:5345-5349.
128. Nobuhisa I, Takeya R, Ogura K, Ueno N, Kohda D, Inagaki F, Sumimoto H: **Activation of the superoxide-producing phagocyte NADPH oxidase requires co-operation between the tandem SH3 domains of p47phox in recognition of a polyproline type II helix and an adjacent alpha-helix of p22phox.** *Biochemical Journal* 2006, **396**(1):183-192.
129. Edens WA, Sharling L, Cheng G, Shapira R, Kinkade JM, Edens HA, Tang X, Flaherty DB, Benian G, Lambeth JD: **Tyrosine cross-linking of extracellular matrix is catalyzed by Duox, a multidomain oxidase/peroxidase with homology to the phagocyte oxidase subunit gp91phox.** *Journal of Cell Biology* 2001, **154**:879-891.
130. Ha EM, Oh CT, Bae YS, Lee WJ: **A direct role for dual oxidase in Drosophila gut immunity.** *Science* 2005, **310**(5749):847-850.
131. Ritsick DR, Edens WA, Finnerty V, Lambeth JD: **Nox regulation of smooth muscle contraction.** *Free Radical Biology & Medicine* 2007, **43**(1):31-38.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

